

Biphasic dose responses in low level light therapy

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Disclosures

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Laser Hair Therapy of North America, Palomar Medical Technologies

Scientific advisory board: Lexington International, Immunophotonics

Outline

- ❖ Introduction to mechanisms of LLLT
- ❖ Survey of biphasic dose responses in LLLT
- ❖ Mechanistic studies in mouse embryonic fibroblasts
- ❖ NF- κ B activation in HEK293 cells
- ❖ HeLa cells and neurons
- ❖ Conclusions

What's in a name?

Low level laser therapy

Low reactive-level laser therapy

Low intensity laser therapy

Low level light therapy

Low energy laser irradiation

Photobiomodulation

Photobiostimulation

Biomodulation

Biostimulation

Cold laser

Soft laser

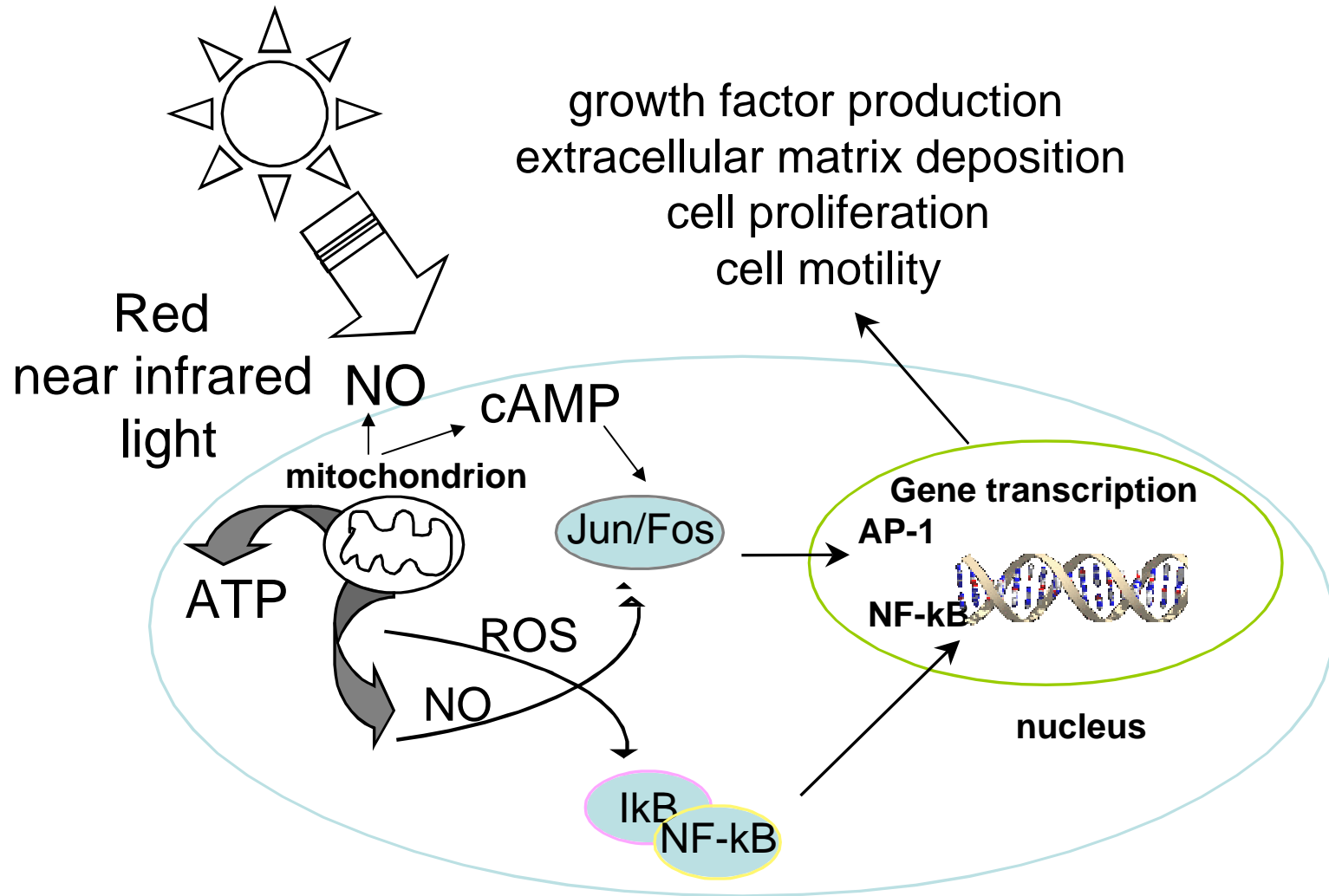
Laser therapy

Phototherapy

It is called “LOW”
because a little
light is better than
a lot of light

Biphasic dose
response?

Mechanisms of LLLT



POWER GAMES

There's a fight going on inside all our cells for each breath of air. **Nick Lane** sheds therapeutic light on the implications for cancer and degenerative diseases.

"The finding that the body could poison one of its own enzymes was initially shrugged off as an imperfection."

Yet over the past decade, researchers have come to appreciate that cells often use CO₂, and to an even greater extent NO (nitric oxide), to block respiration. Not only that, but light has striking counter-effects on cytochrome oxidase. And all these suitors to the enzyme turn out to be critical to our understanding not just

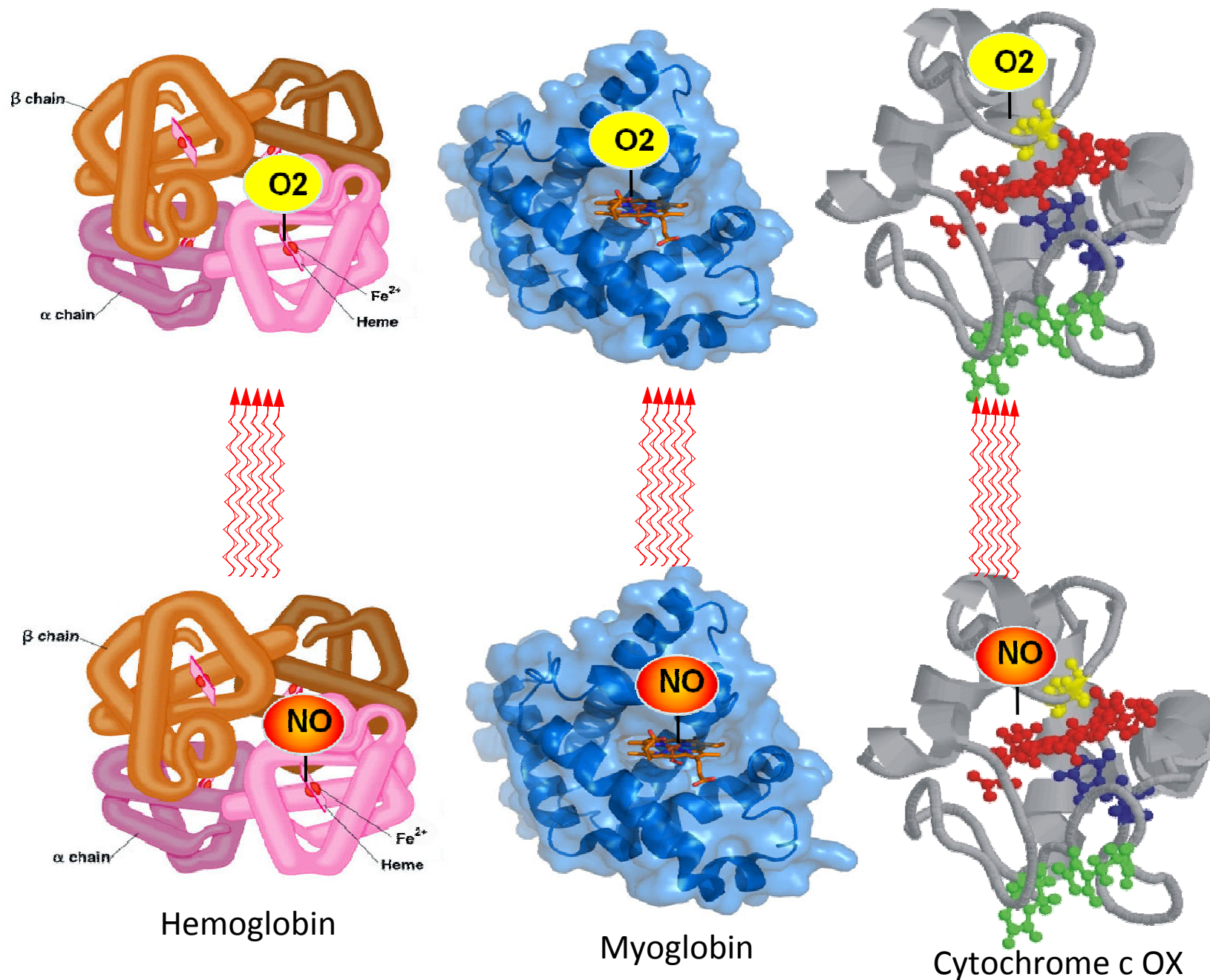
Nitric Oxide and the Control of Firefly Flashing

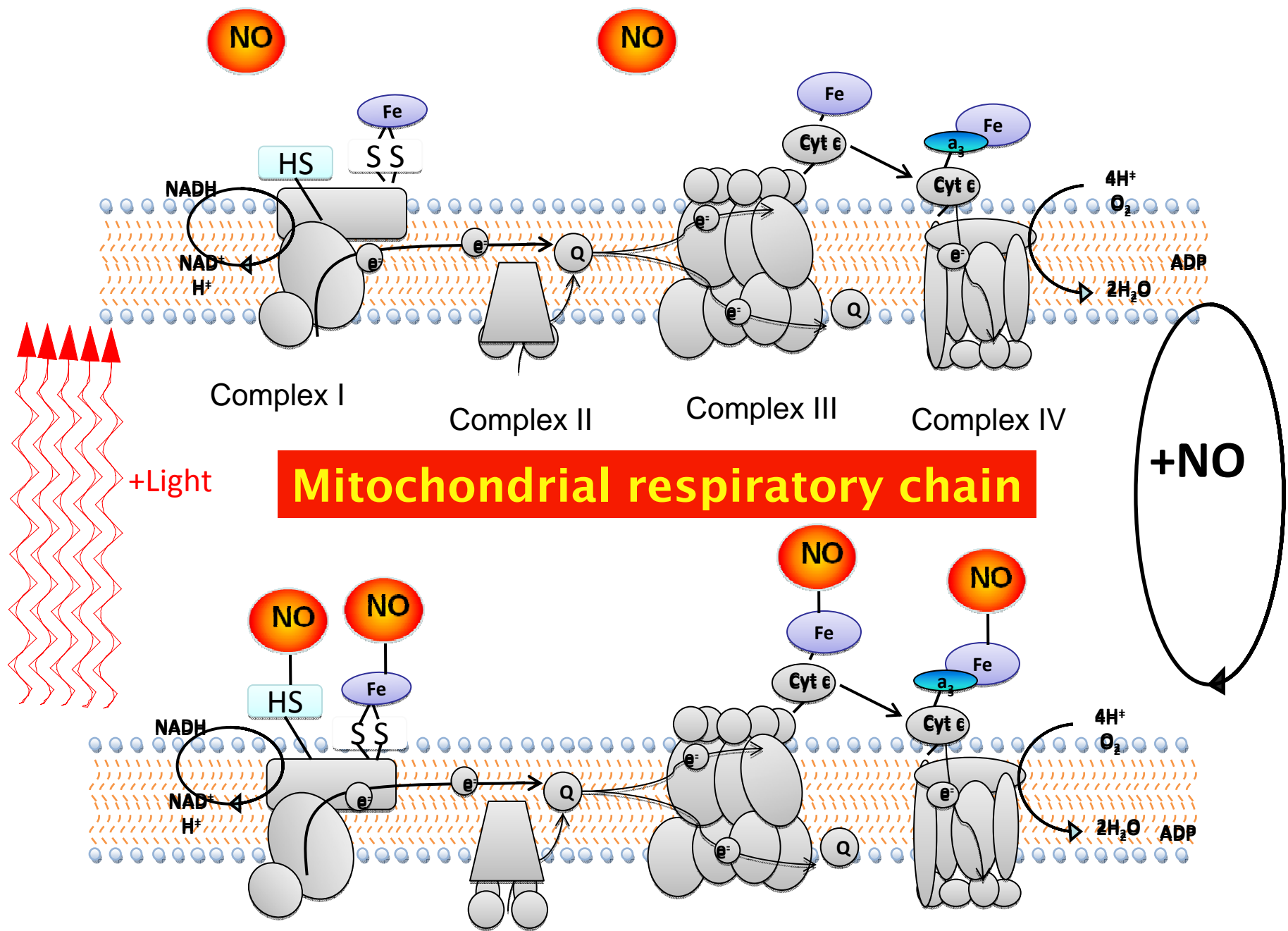
Barry A. Trimmer,^{1*} June R. Aprille,¹ David M. Dudzinski,²
Christopher J. Lagace,¹ Sara M. Lewis,¹ Thomas Michel,^{2,3}
Sanjive Qazi,¹ Ricardo M. Zayas¹

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29 JUNE 2001 VOL 292 SCIENCE

The results reported here document an important role for NO in firefly flash control. It is well established that O₂ availability is the immediate biochemical trigger for light production, and we propose that the role of NO is to transiently inhibit mitochondrial respiration in photocytes and thereby increase O₂ levels in the peroxisomes. This is consistent with the distinctive spatial arrangement of NOS-containing cells, the known NO-mediated inhibition of cytochrome c oxidase (21–23), and the fact that firefly luminescence can be induced by cytochrome c oxidase inhibitors, such as cyanide and carbon monoxide





Cytochrome c oxidase can act as a photoreceptor allowing the photolytic dissociation of any bound nitric oxide.

Karu, T.I., L.V. Pyatibrat, and G.S. Kalendo, Photobiological modulation of cell attachment via cytochrome c oxidase. Photochem Photobiol Sci, 2004. 3(2): p. 211-6.

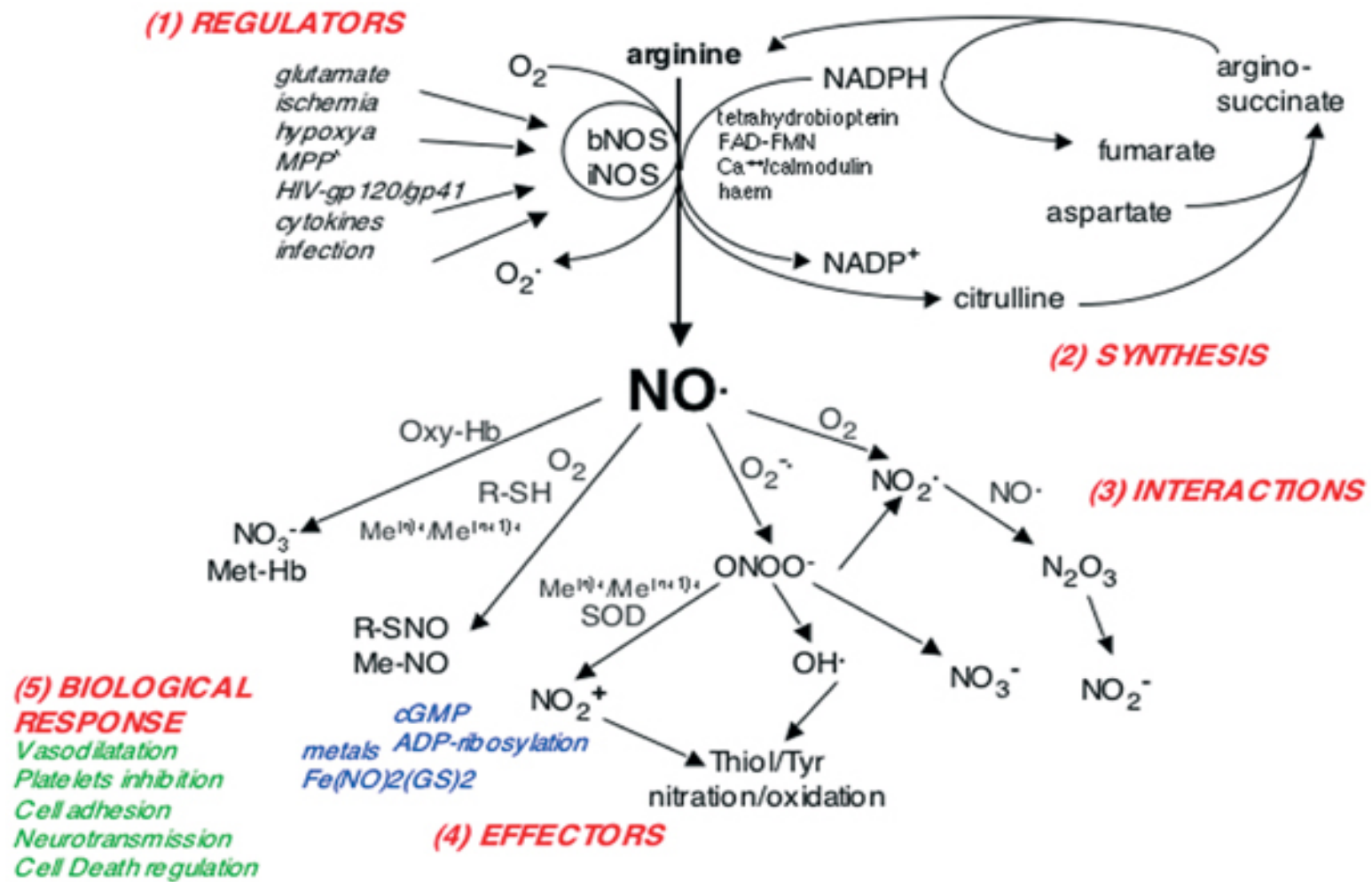
Karu, T.I., et al., Absorption measurements of a cell monolayer relevant to phototherapy: reduction of cytochrome c oxidase under near IR radiation. J Photochem Photobiol B, 2005. 81(2): p. 98-106.

The nitric oxide present in the heme-CuLa3 center of CytC ox can be photolysed by visible light.

Sarti, P., et al., Nitric oxide and cytochrome c oxidase: mechanisms of inhibition and NO degradation. Biochem Biophys Res Commun, 2000. 274(1):p. 183-7.

Near infrared light protects cardiomyocytes from hypoxia and reoxygenation injury by a nitric oxide dependent mechanism.

Zhang et al. Journal of Molecular and Cellular Cardiology 46 (2009) 4–14



Basic mechanisms

Mitochondria are primary photoreceptors

Cytochrome c oxidase activity is increased

NO is dissociated from COX + heme proteins

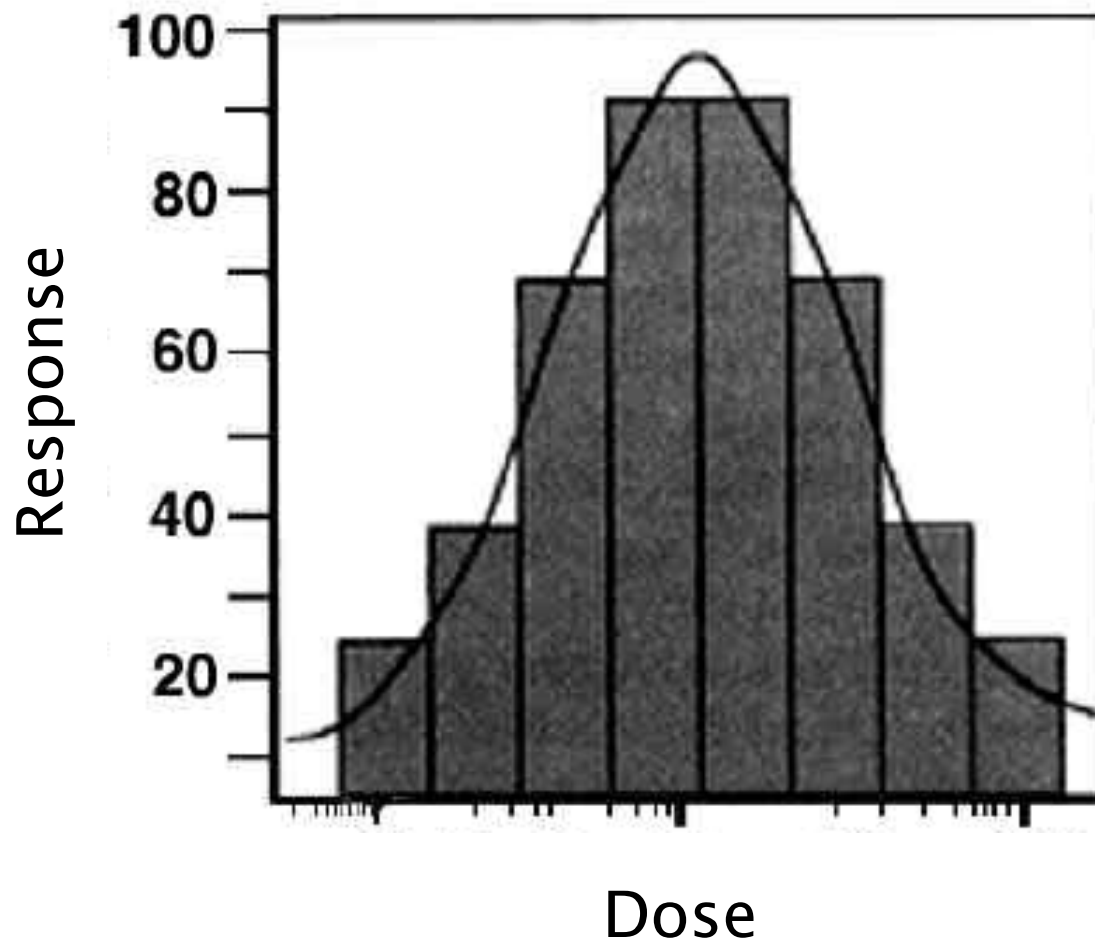
ATP and cAMP increased

Reactive oxygen species are produced

Transcription factor induction

Growth, repair, survival, less inflammation

Biphasic dose response?



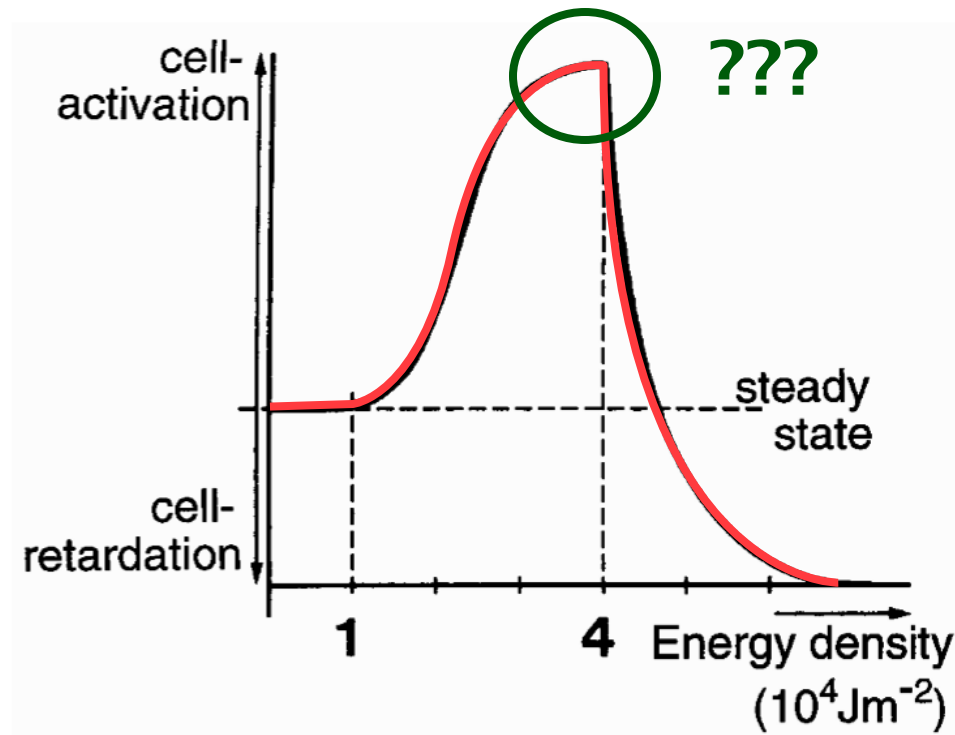
How is dose measured?

Power (W) x Time (sec) = Energy (Joules)

$$\frac{\text{Power mW}}{\text{Beam Area cm}^2} = (\text{irradiance}) \times \text{time} = \text{fluence (J/cm}^2\text{)}$$

Arguments have been made for total energy, fluence, irradiance and illumination time to be most important parameters in measuring dose

- Dose response curve

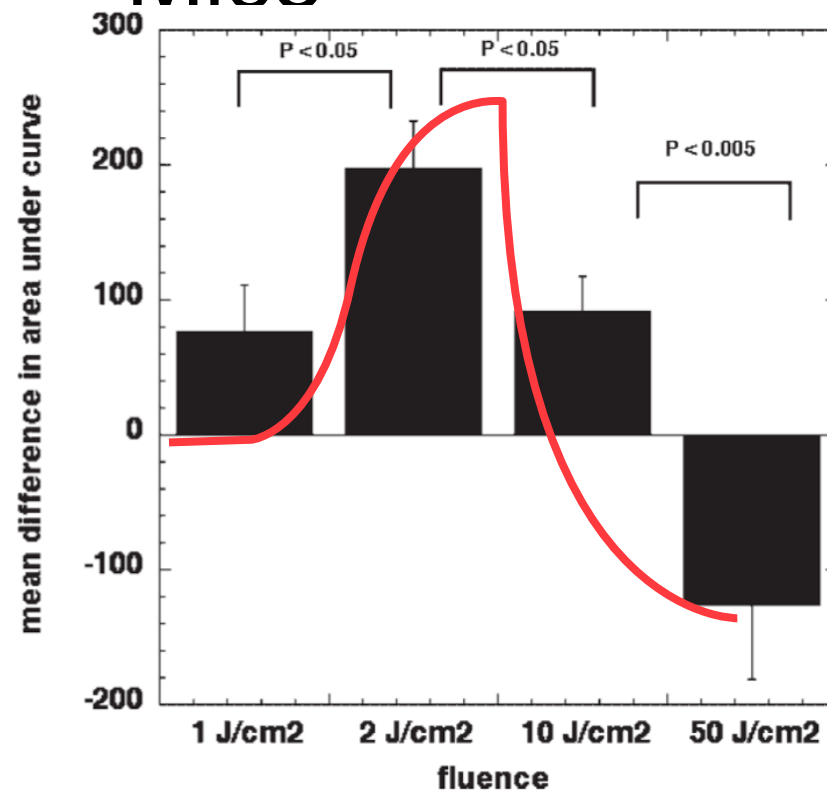


Sommer AP, Pinheiro AL, Mester AR, Franke RP, Whelan HT. (2001) Biostimulatory windows in low-intensity laser activation: lasers, scanners, and NASA's light-emitting diode array system. J Clin Laser Med Surg. Feb;19(1):29-33

Arndt-Schulz curve

Stimulation/Inhibition of Wound Healing in Mice

635-nm Laser for wound healing in mice



Lasers in Surgery and Medicine 39:706–715 (2007)

Low-Level Light Stimulates Excisional Wound Healing in Mice

Tatiana N. Demidova-Rice, BS,^{1,2} Elena V. Salomatina, BS,¹ Anna N. Yaroslavsky, PhD,^{1,3} Ira M. Herman, PhD,^{2*} and Michael R. Hamblin, PhD^{1,3,4**}

¹Wellman Center for Photomedicine, Massachusetts General Hospital, Boston, Massachusetts 02114

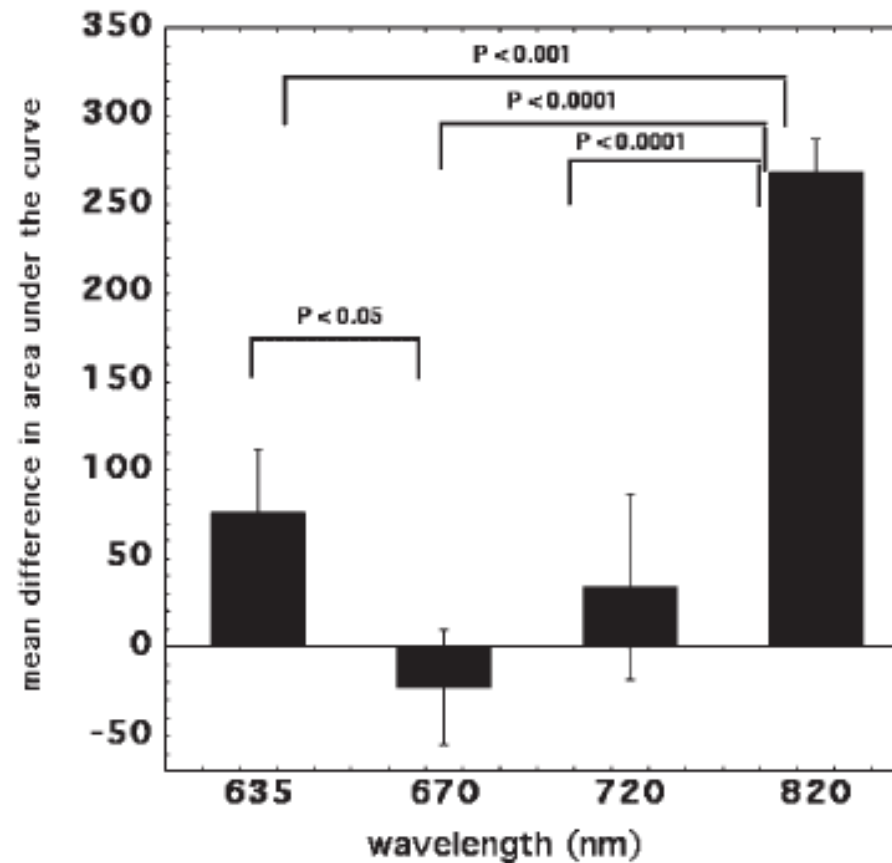
²Graduate Program in Cell Molecular and Developmental Biology, Sackler School of Graduate Biomedical Sciences, Tufts University School of Medicine, Boston, Massachusetts 02111

³Department of Dermatology, Harvard Medical School, Boston, Massachusetts 02115

⁴Harvard-MIT Division of Health Sciences and Technology, Cambridge, Massachusetts 02139

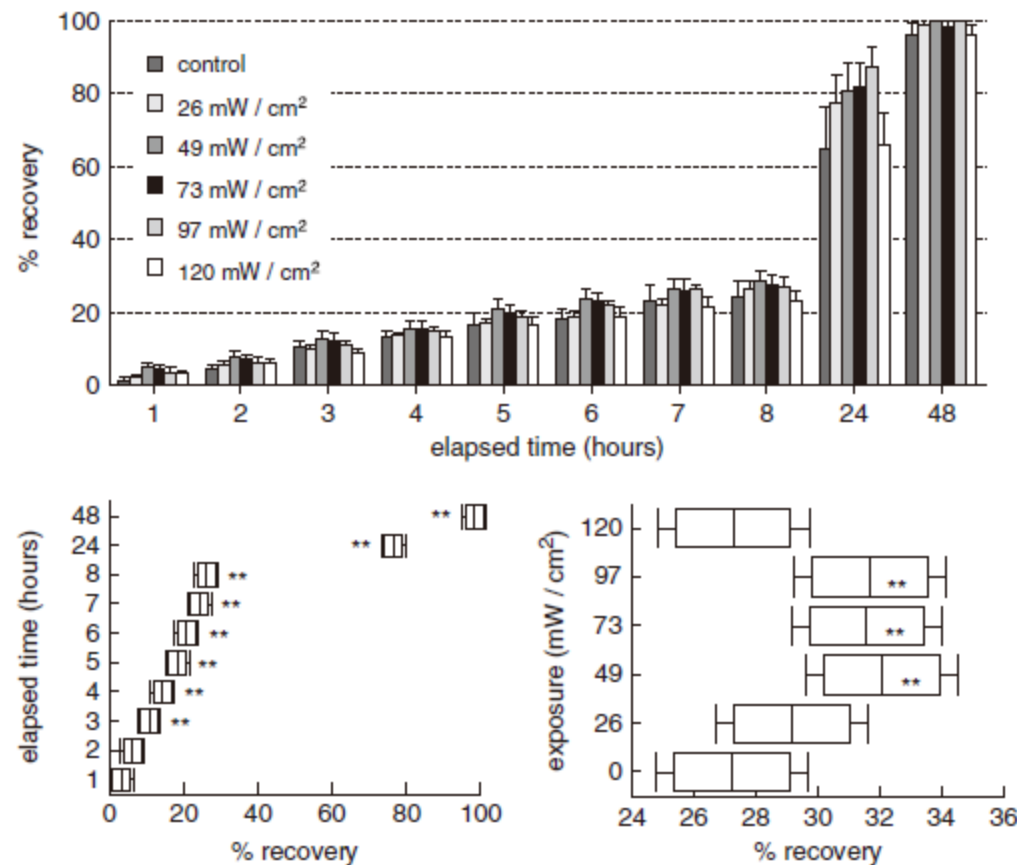
Michael R Hamblin, PhD, Wellman Center for Photomedicine. Dose Response Conference 2009

Wavelength response (action spectrum?)



Biphasic dose response may be different at each wavelength

Wound healing in vitro 980-nm laser - scratch in fibroblast monolayer



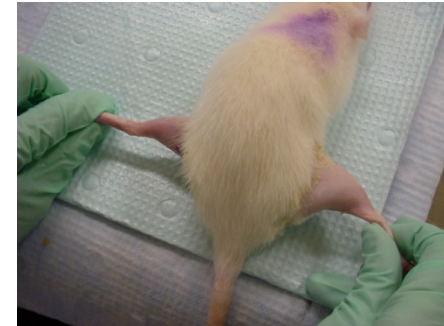
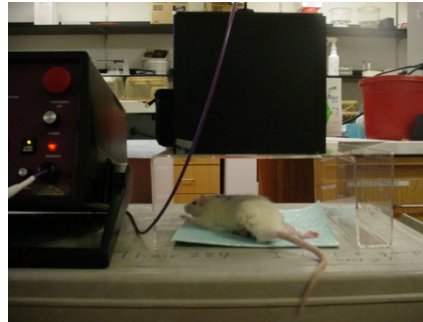
Constant time - irradiance and fluence vary

Mark D. Skopin & Scott C. Molitor

Effects of near-infrared laser exposure in a cellular model of wound healing

Photodermatol Photoimmunol Photomed, **25**, 75-80.(2009).

810-nm laser for arthritis in rats



| | 3 J/cm ² | 30 J/cm ² |
|-----------------------|---------------------|----------------------|
| 5 mW/cm ² | 10 minutes | 100 minutes |
| 50 mW/cm ² | 1 minute | 10 minutes |

Hypothesis: length of illumination is more important than total fluence or irradiance in LLLT effect

Lasers in Surgery and Medicine 39:543–550 (2007)

Low-Level Laser Therapy for Zymosan-Induced Arthritis in Rats: Importance of Illumination Time

Ana P. Castano, MD,^{1,2} Tianhong Dai, PhD,^{1,2} Ilya Yaroslavsky, PhD,³ Richard Cohen, MD,³ William A. Apruzzese, PhD,³ Michael H. Smotrich, PhD,³ and Michael R. Hamblin, PhD^{1,2,4*}

¹Wellman Center for Photomedicine, Massachusetts General Hospital, Boston, Massachusetts 02114

²Department of Dermatology, Harvard Medical School, Boston, Massachusetts 02115

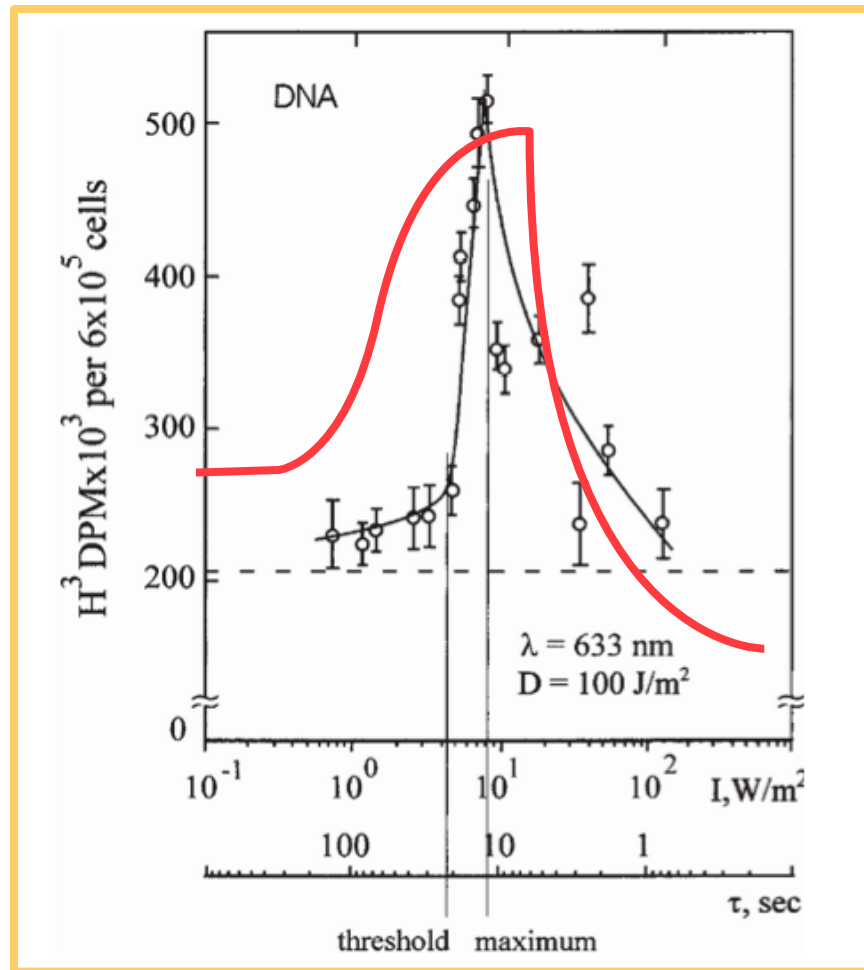
³Palomar Medical Technologies Inc., Burlington, Massachusetts 01803

⁴Harvard-MIT Division of Health Sciences and Technology, Cambridge, Massachusetts 02139

Michael R Hamblin, PhD, Wellman Center for Photomedicine. Dose Response Conference 2009

- Dose response curve

HeLa DNA synthesis
633nm
0.1J/cm²
10 - 1000mW/cm²



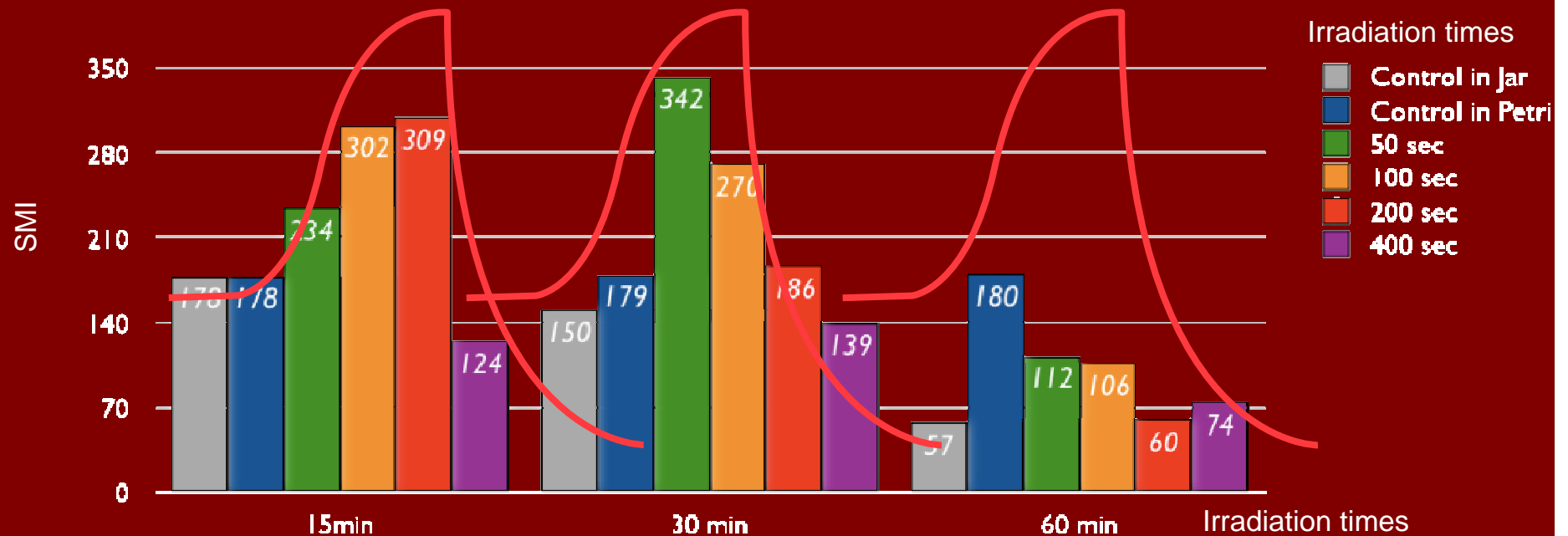
increase power
reduce time
(same energy)

Karu TI, Kolyakov SF. (2005) Exact action spectra for cellular responses relevant to phototherapy. Photomed Laser Surg. Aug;23(4):355-61.

Michael R Hamblin, PhD, Wellman Center for Photomedicine. Dose Response Conference 2009

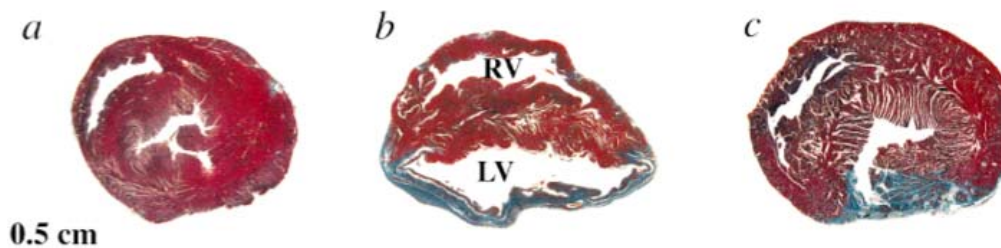
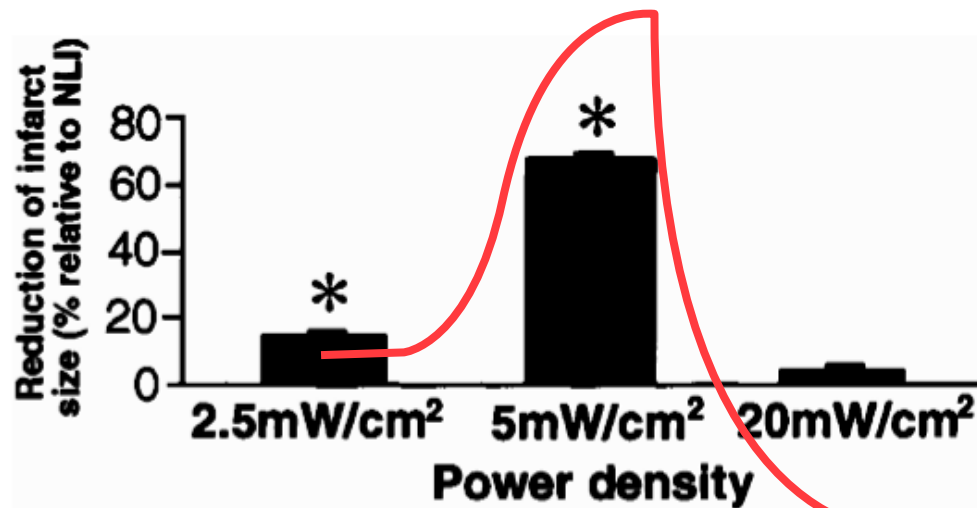


Motility changes over time Fresh Semen LED 45mW/cm²



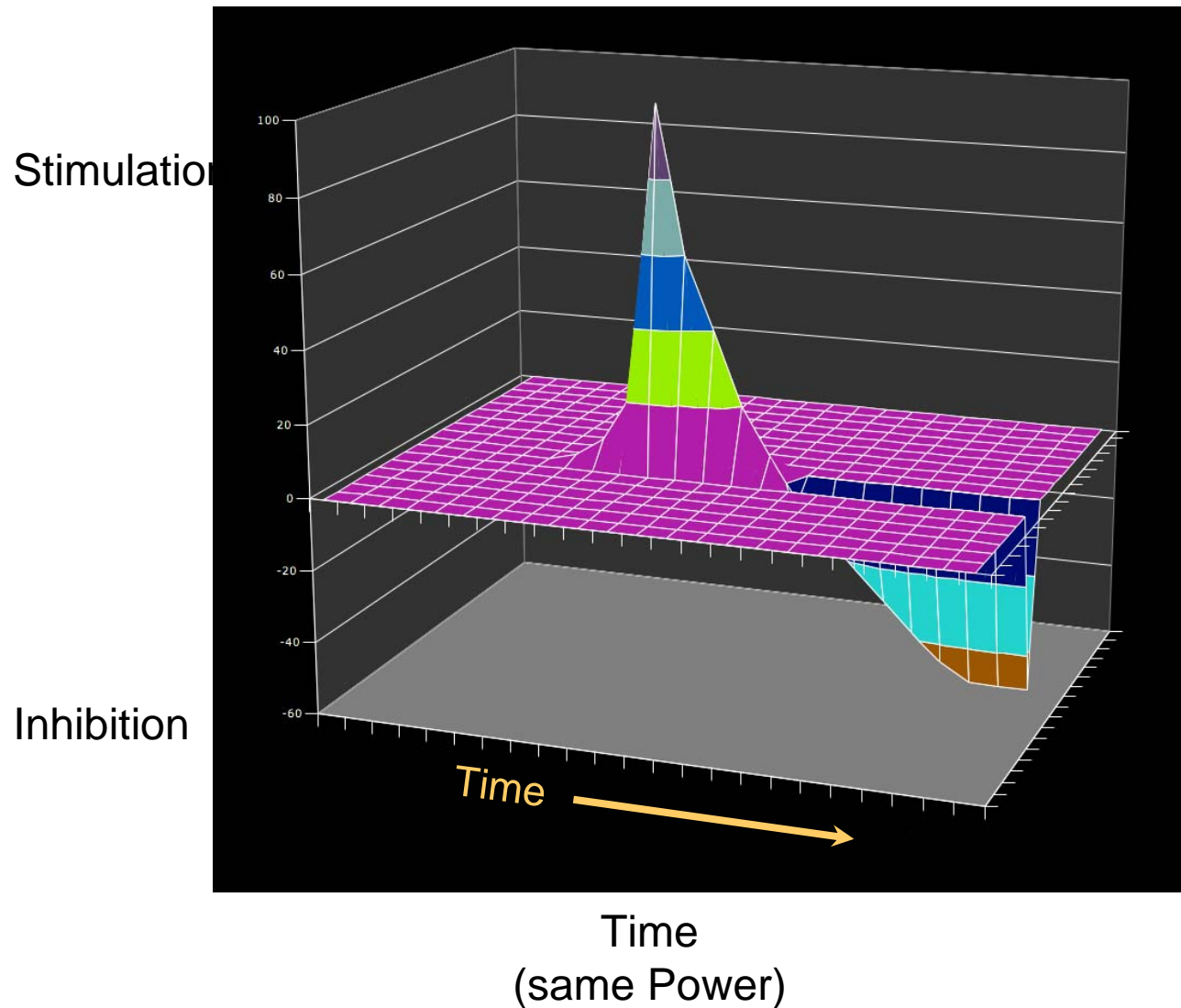
Motility changes over time
with different irradiation times using
104 x LED Cluster mixed 660nm & 850nm
45mW/cm²

Reduction of infarct size (heart attack) by low-energy laser

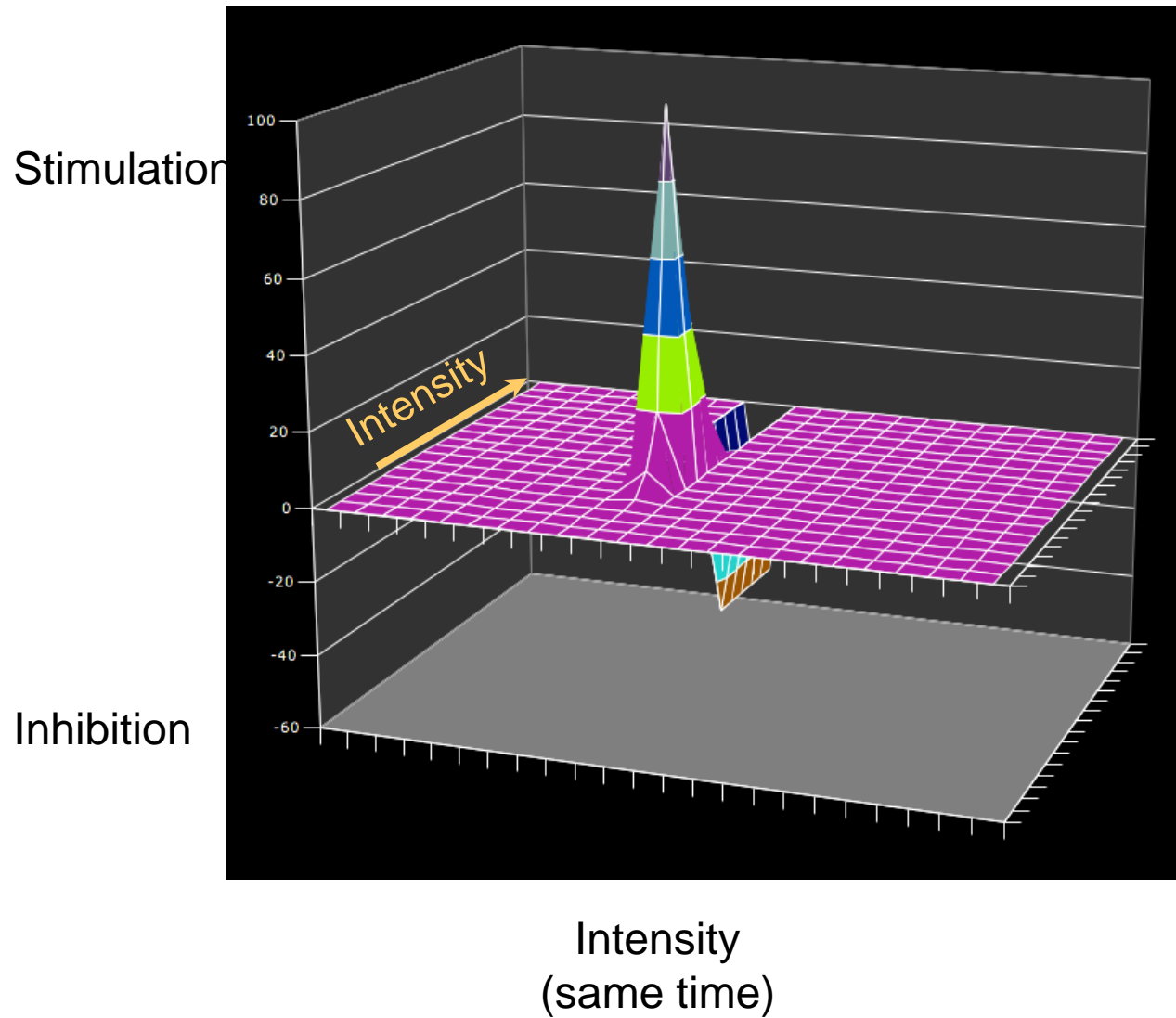


Oron U, et al
Attenuation of infarct size in rats and dogs after
myocardial infarction by low-energy laser
irradiation. Lasers Surg Med.;28(3):204-11.

- Dose response curve



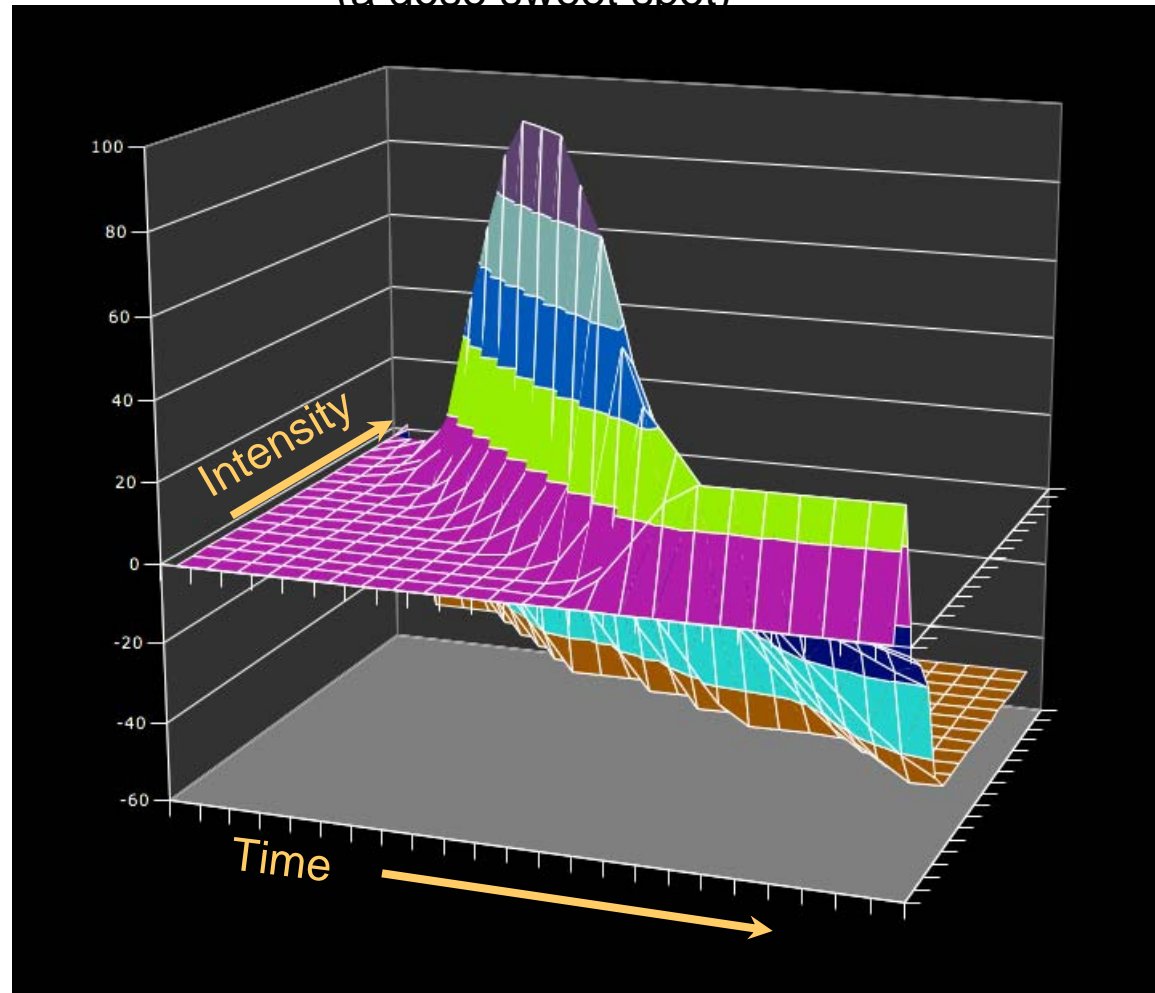
- Dose response curve



A 3D model for Low Level Laser / LED therapy
Biostimulation and Bioinhibition
(a dose sweet spot)

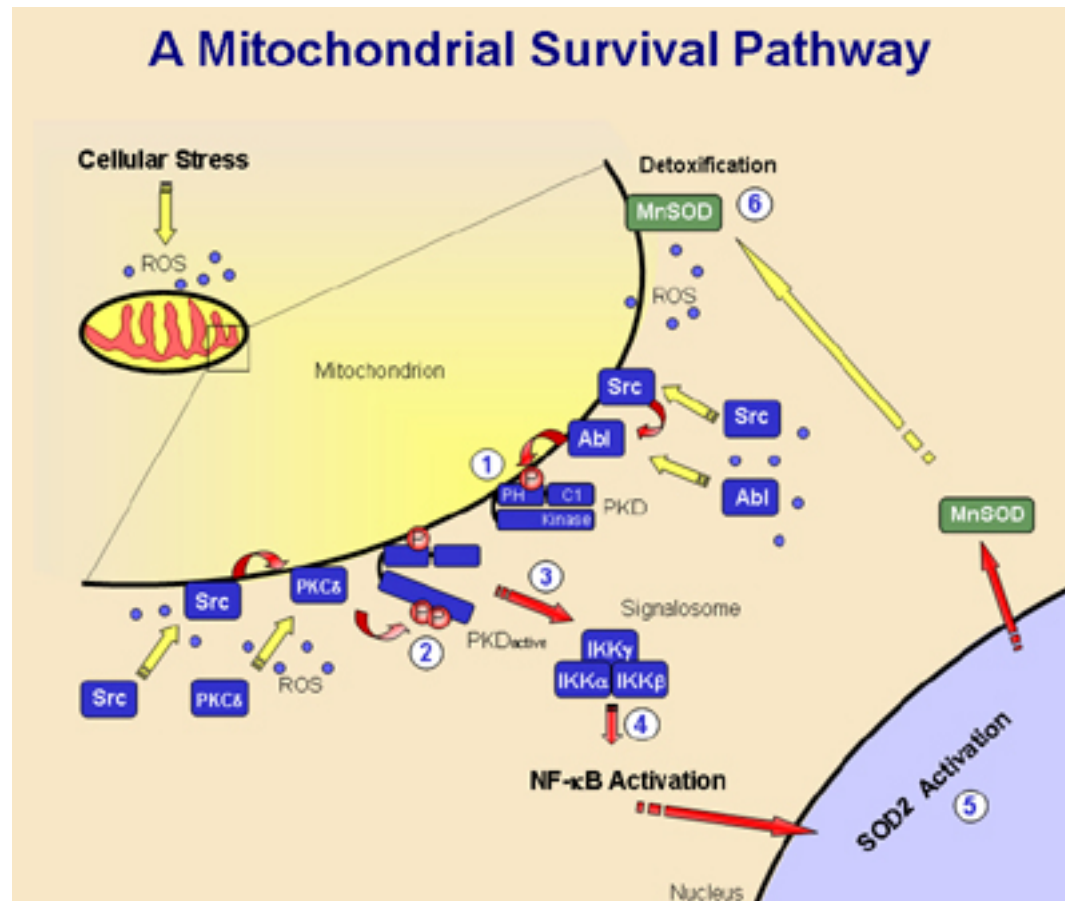
Stimulation

Inhibition

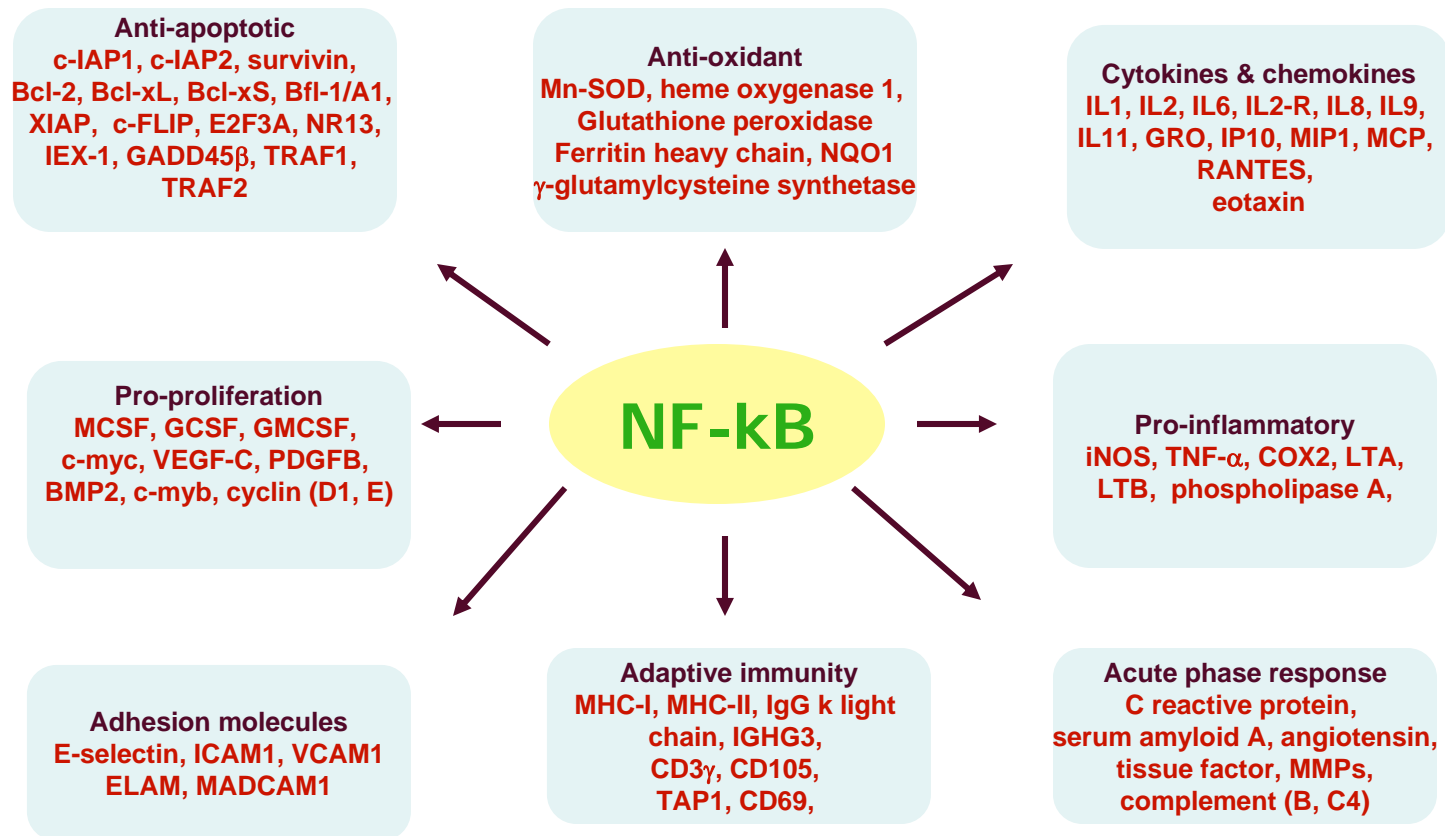


Anders Island

Mitochondrial ROS induces transcription factor NF- κ B



NF-kB target genes

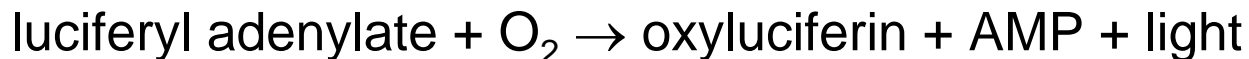


Use of luciferase and bioluminescence

Luciferase is a generic name for enzymes commonly used in nature for bioluminescence.

Firefly luciferase (EC 1.13.12.7) from the firefly *Photinus pyralis*.

Light is produced by the oxidation of luciferin substrate consuming adenosine triphosphate (ATP) and catalyzed by luciferase



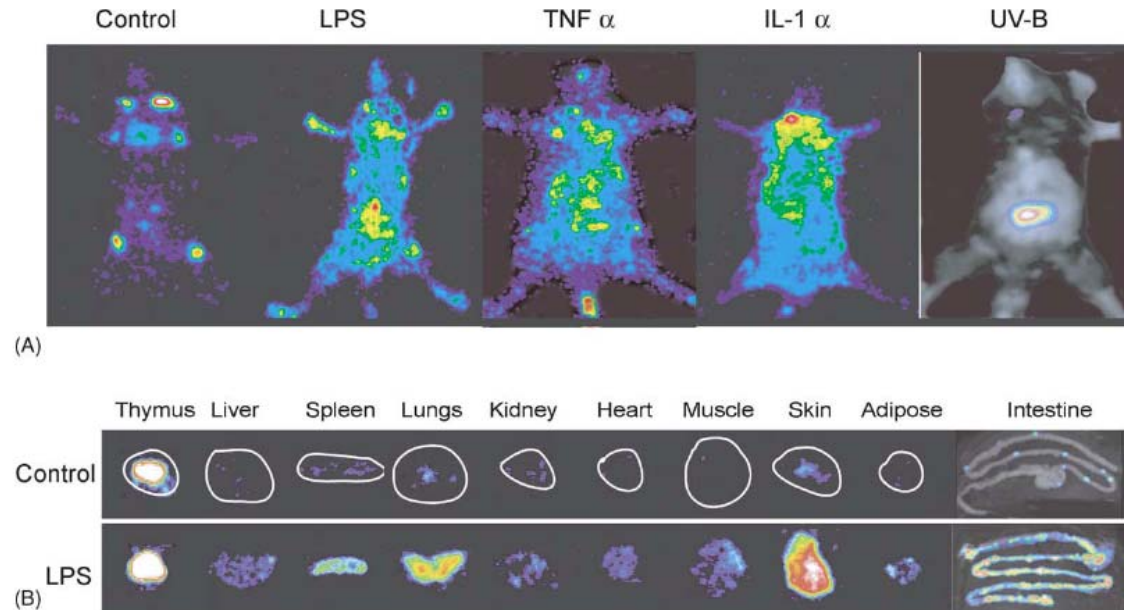
Review

Molecular imaging of the transcription factor NF- κ B, a primary regulator of stress response

Harald Carlsen, George Alexander, Liv M.I. Austenaa,
Kanae Ebihara, Rune Blomhoff*

Department of Nutrition, Faculty of Medicine, University of Oslo, P.O. Box 1046 Blindern, N-0316 Oslo, Norway

Received 3 October 2003; received in revised form 23 February 2004; accepted 23 February 2004



NF-kB response element on Ig-k light chain promoter drives luciferase

Does LLLT activate NF-kB?

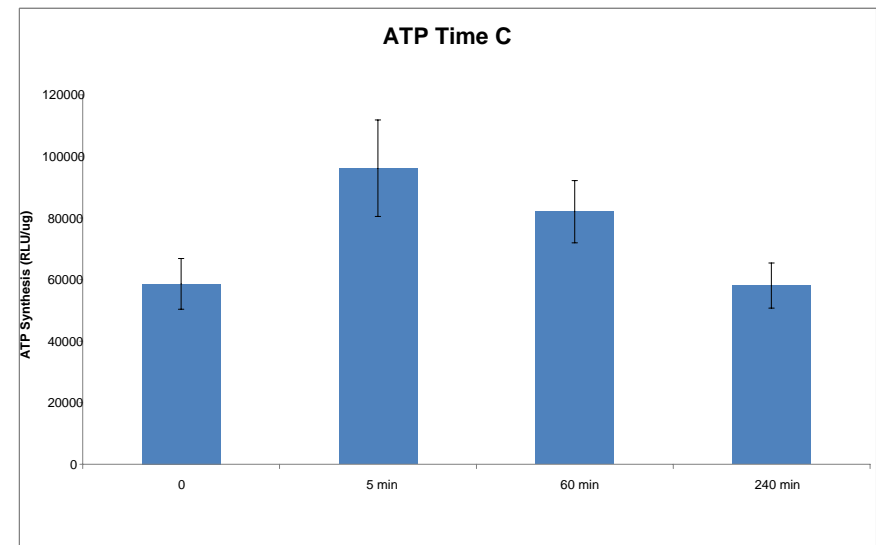
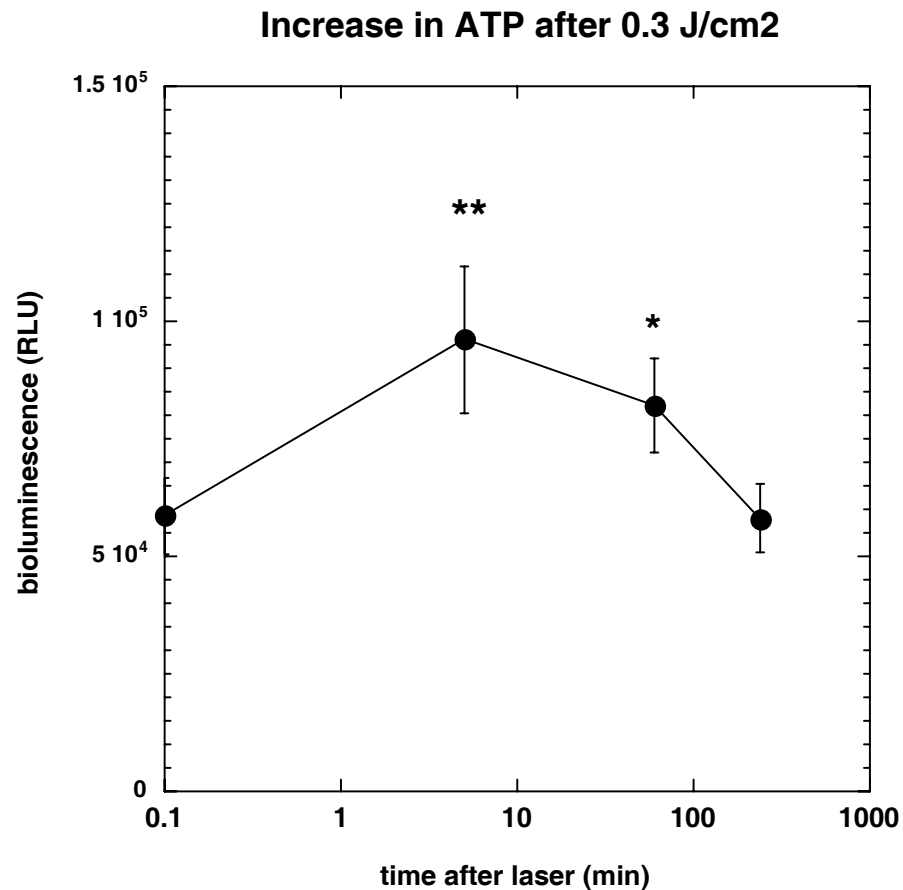
1. Establish a fibroblast cell line (3T3 protocol) from NF-kB luciferase reporter mice (HLL)
2. Deliver different fluences of 810-nm light from a laser (or other light source)
3. Keep illumination time constant at 5 min (vary irradiance)
4. After various times assay for luciferase expression (NF-kB activation) and cellular ATP

810-nm laser

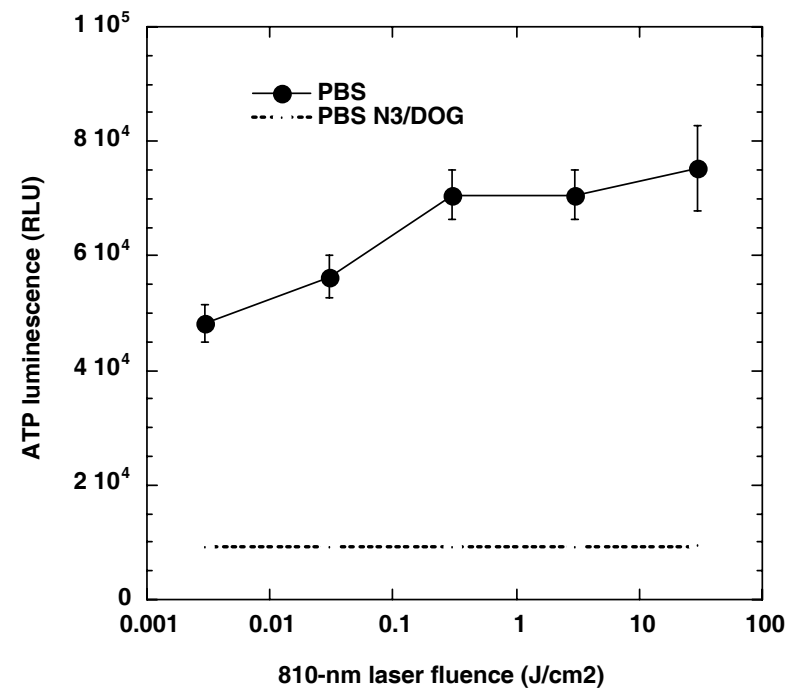
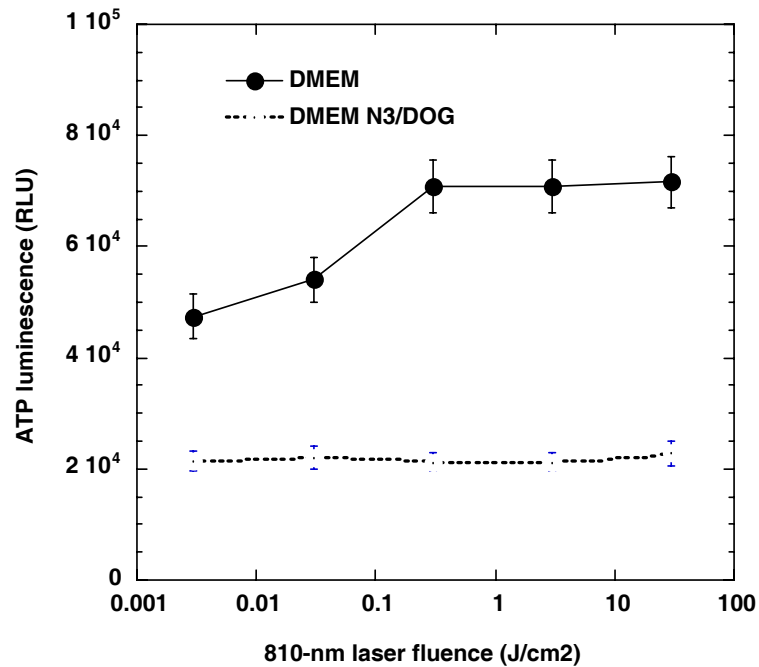


1. Effects on ATP production

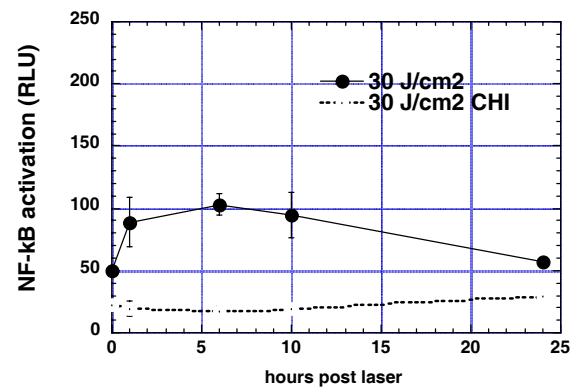
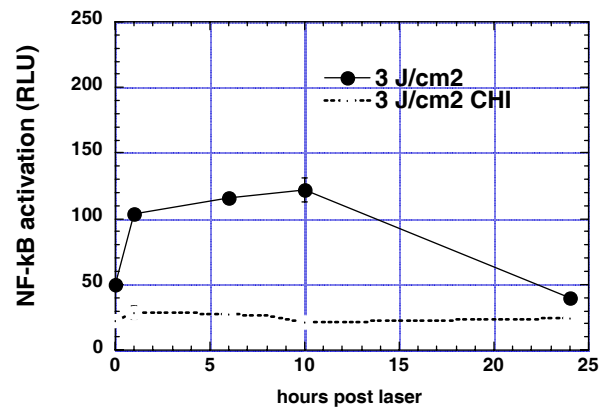
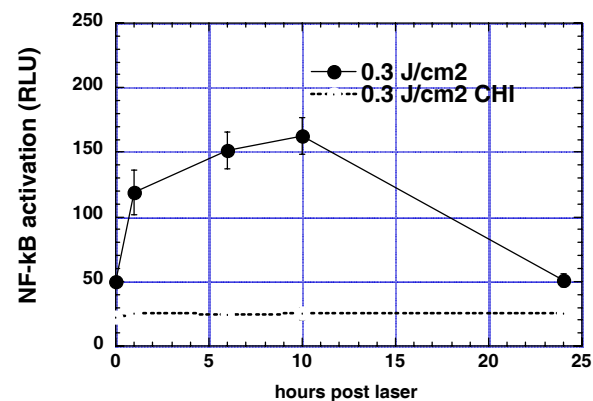
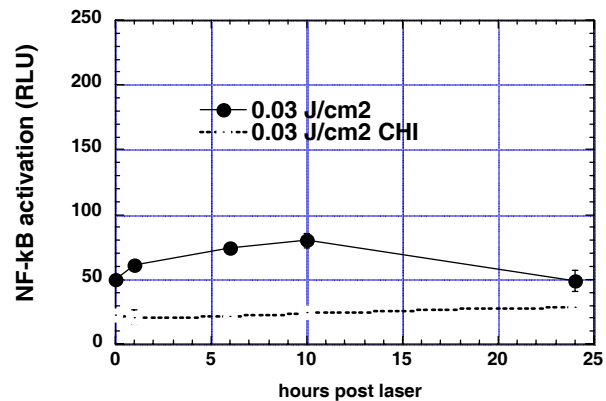
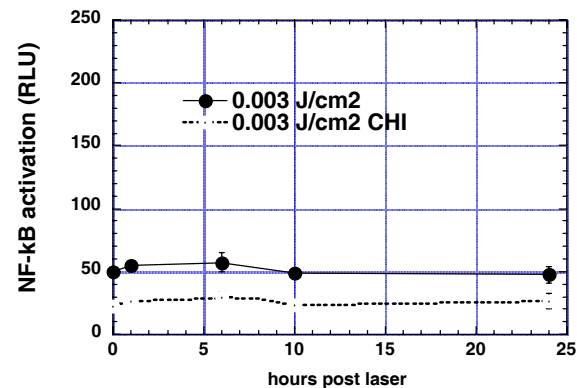
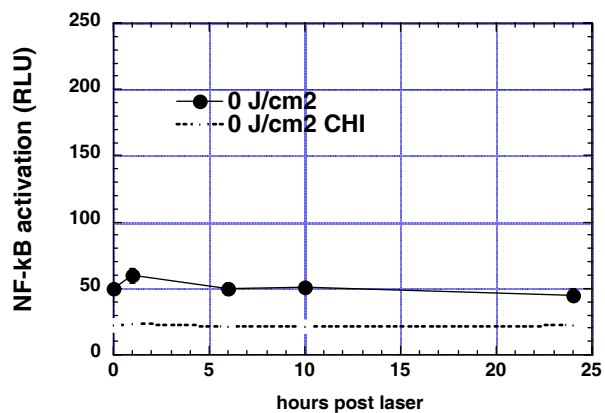
0.3 J/cm² 810-nm delivered over 5 min



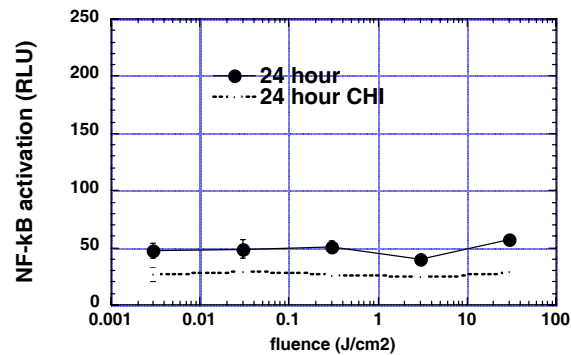
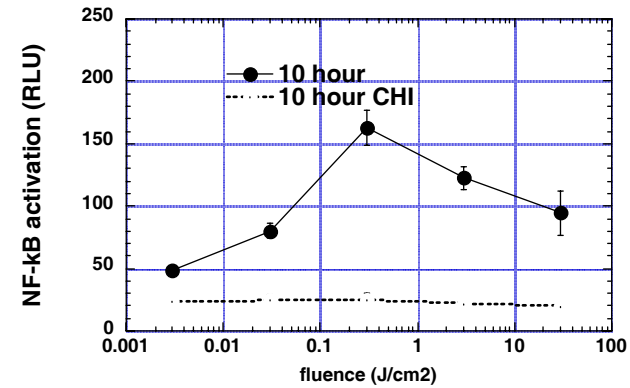
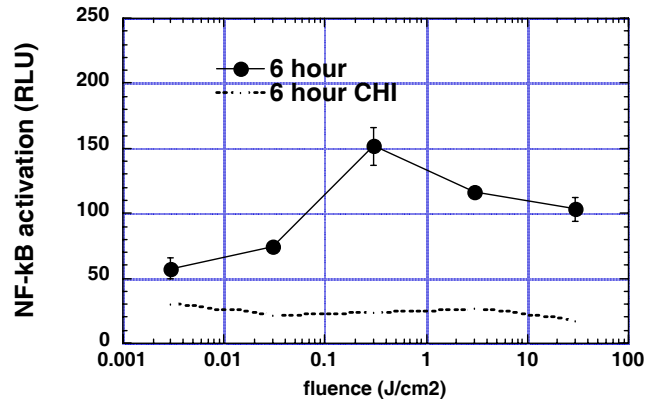
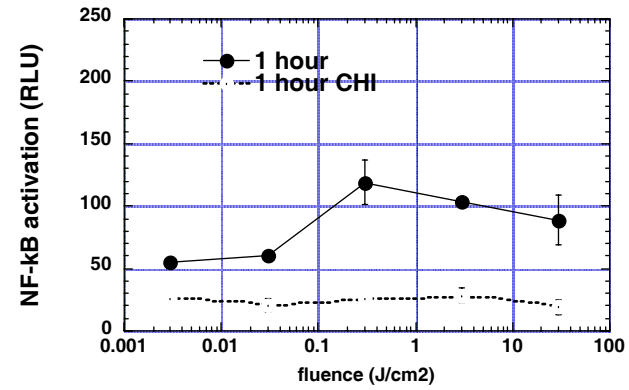
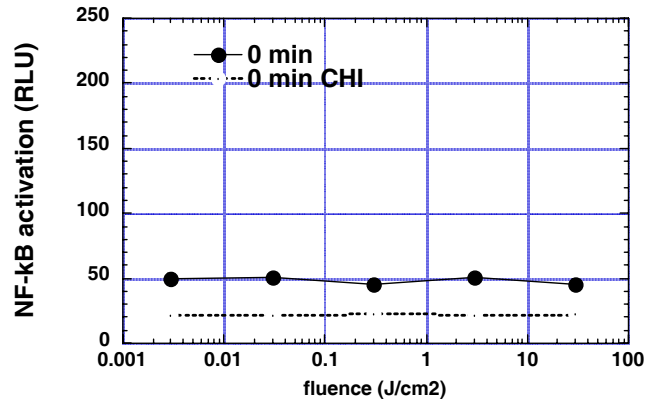
Fluence effect (dose-response) of ATP increase (measured at 5 min post laser) Azide/deoxyglucose mimics hypoxia 0.003, 0.03, 0.3, 3 and 30 J/cm²



2. Time course of NF-kB activation

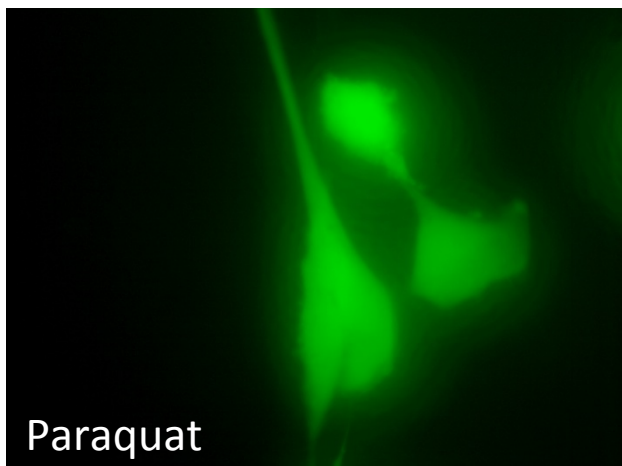
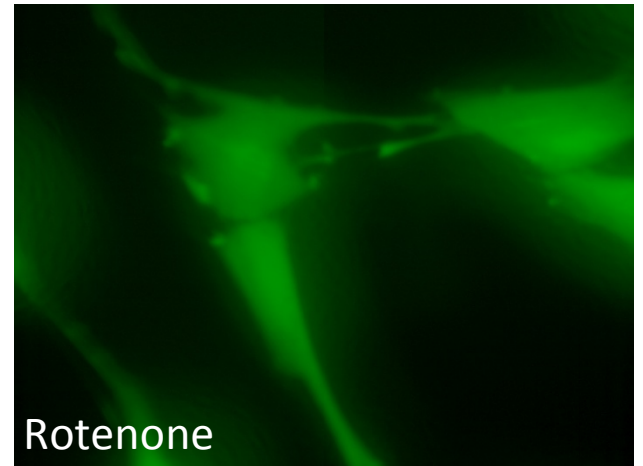
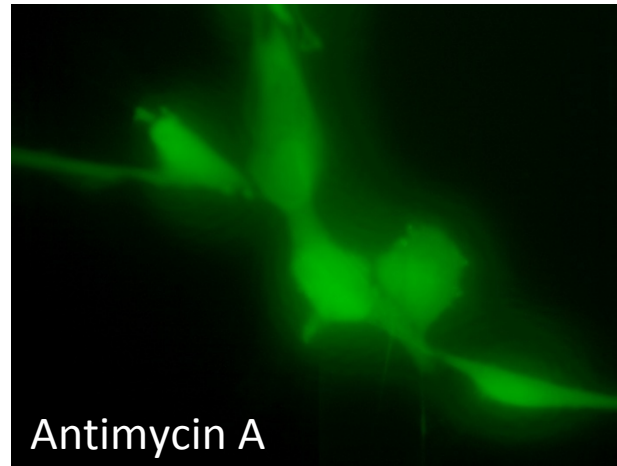
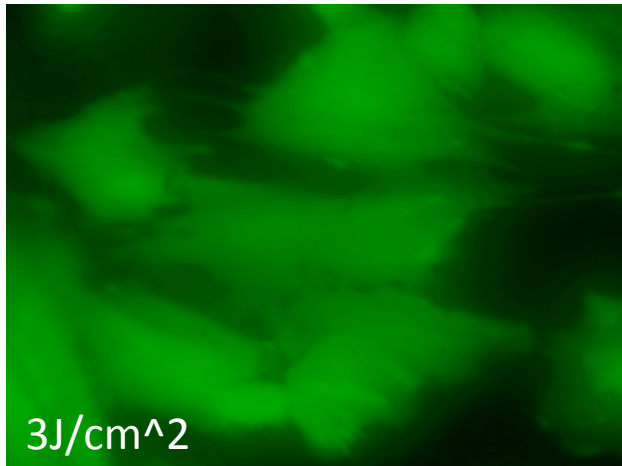
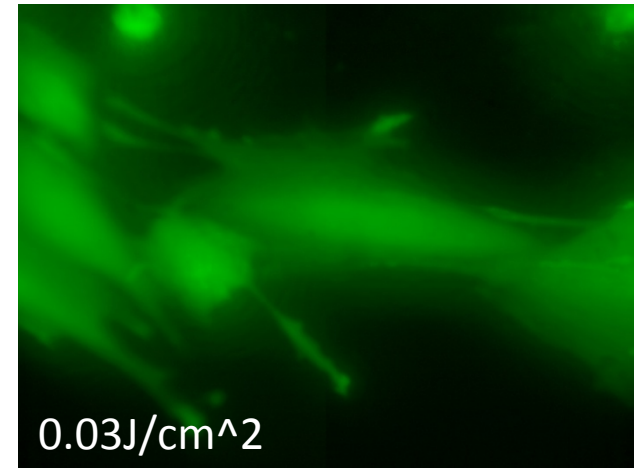
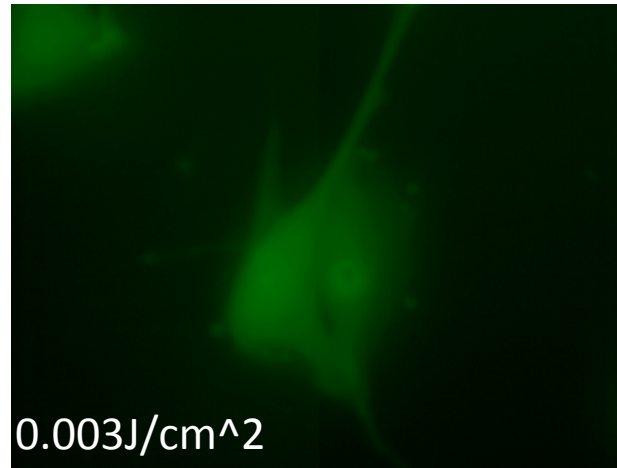
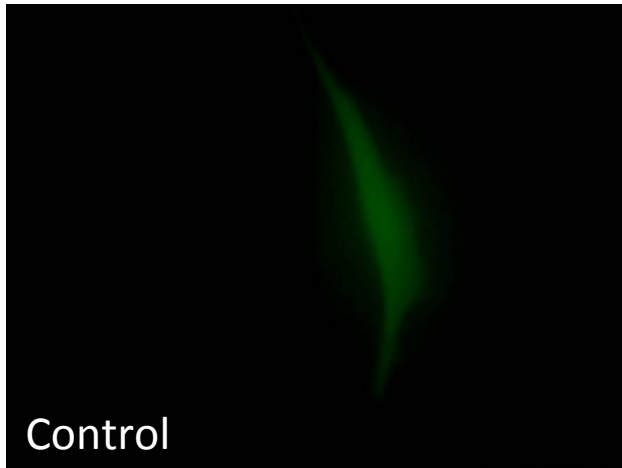


Fluence response of NF-kB activation



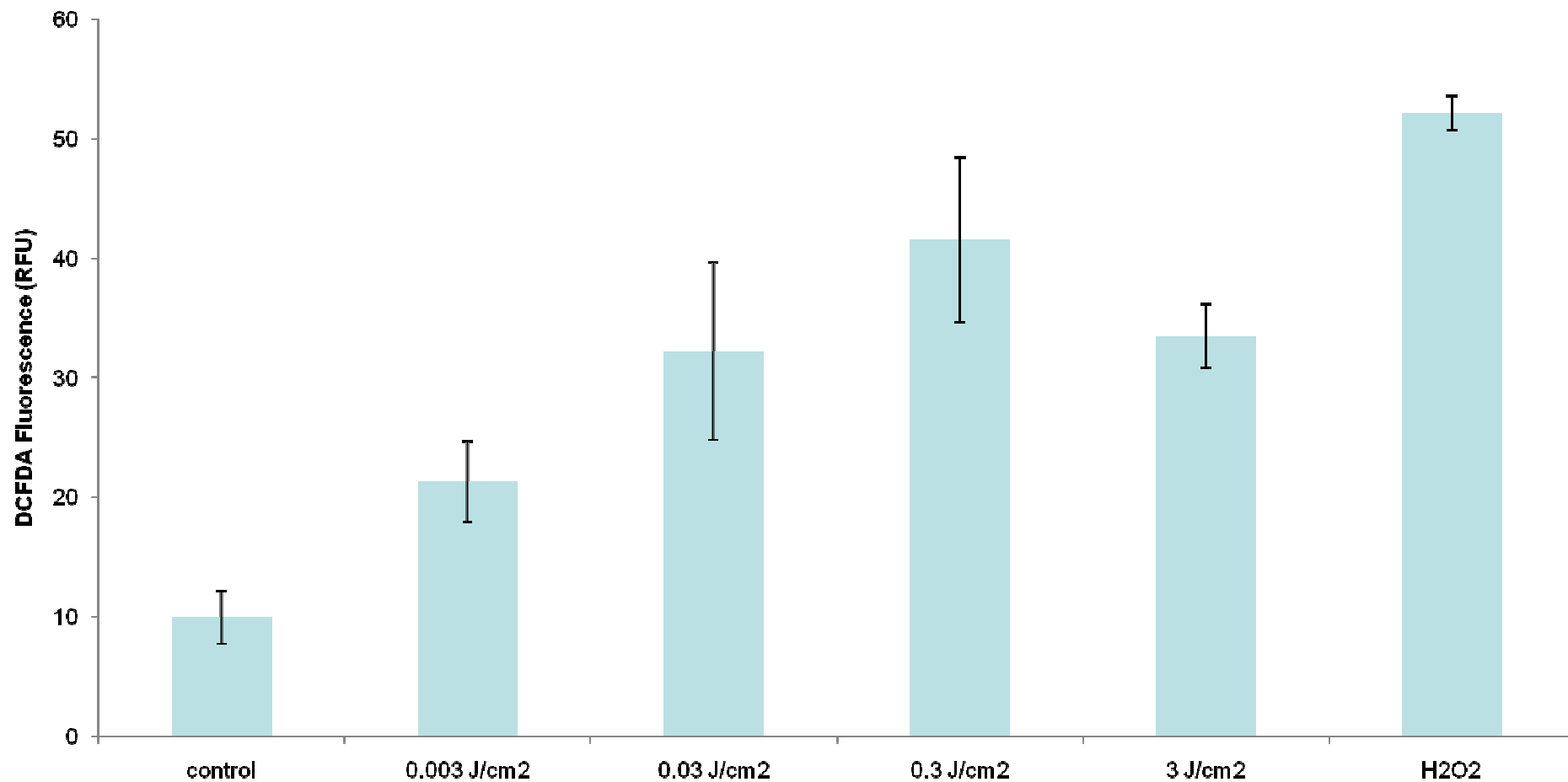
What is mechanism of NF-kB activation?

Hypothesis is reactive oxygen species



DCFDA
Sensitive to lipid hydroperoxides

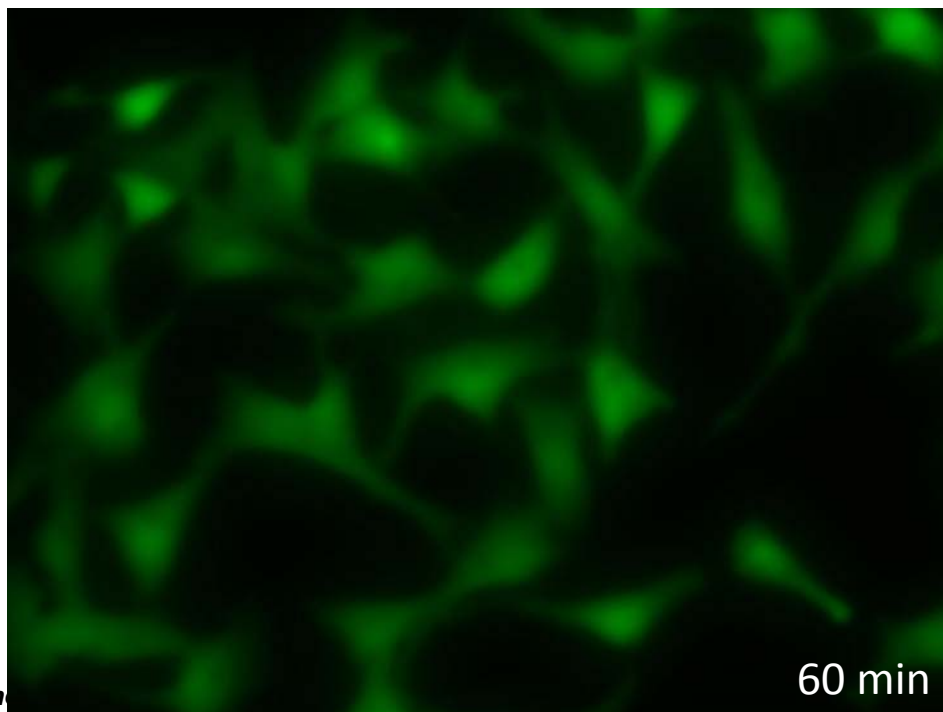
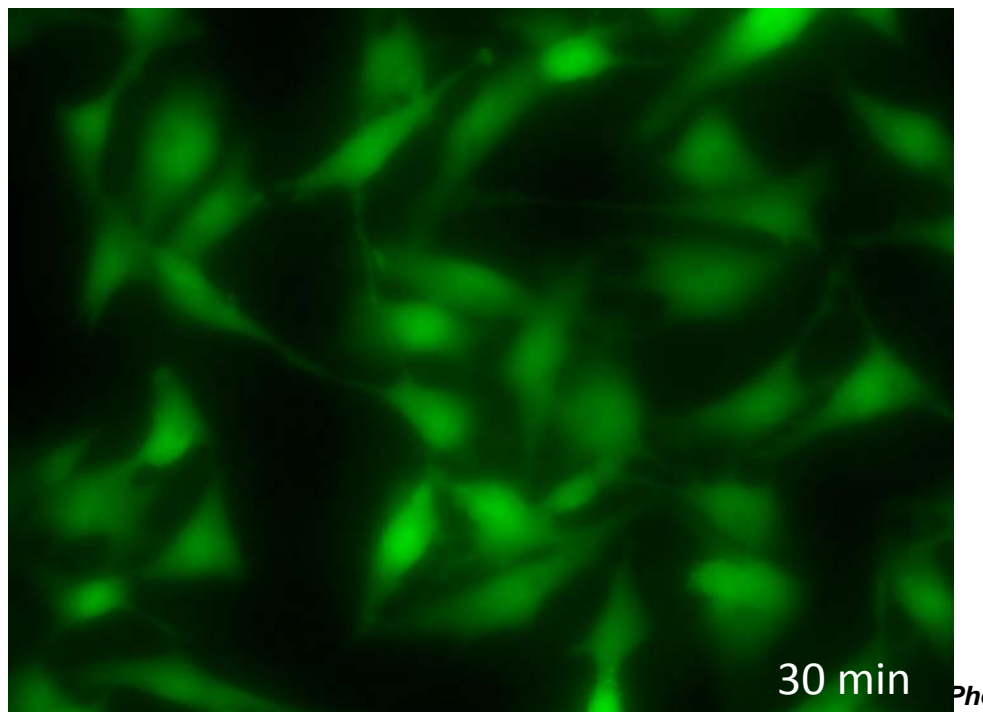
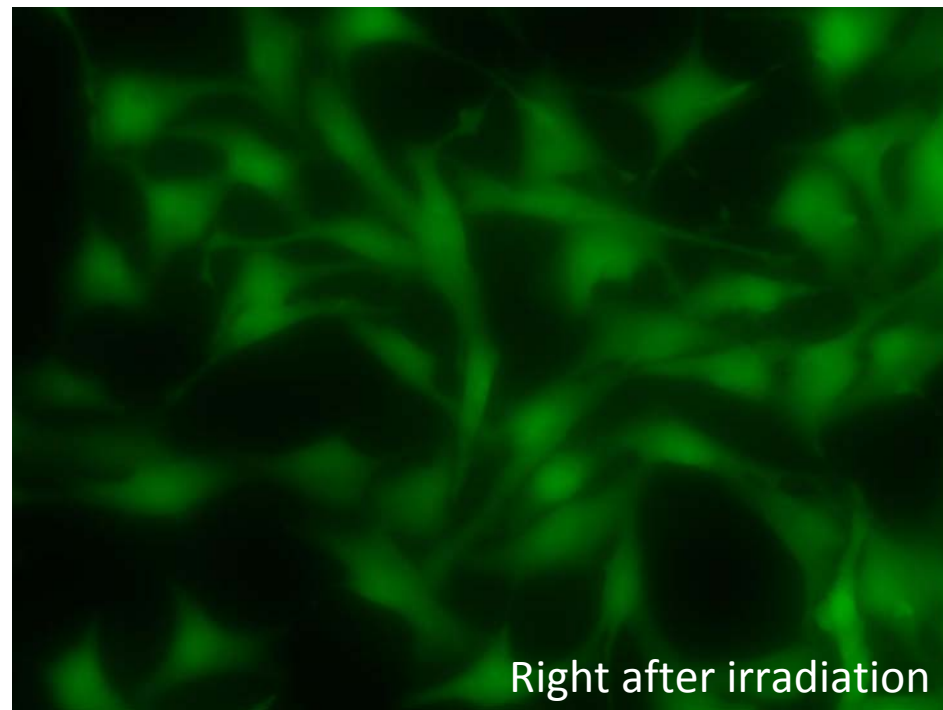
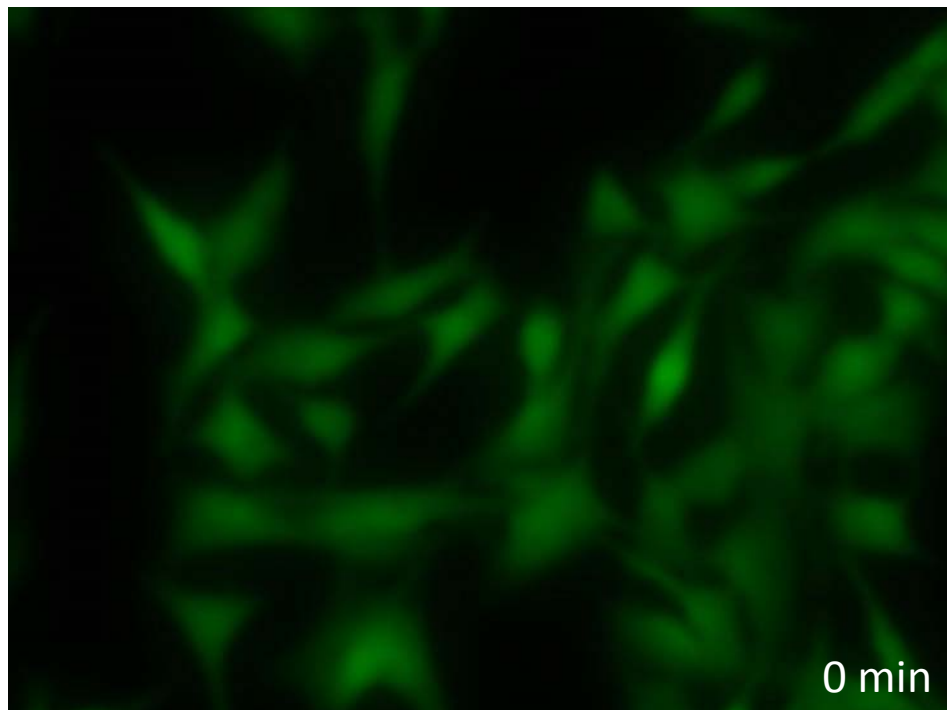
Quantification of ROS induced by laser



MEF ROS Time Course

MEF ROS Time course

- 1. Irradiate MEF with 810nm Laser
- 2. At each time point, 2x washing with PBS and add the DCFDA probe.
- 3. Incubate the MEF with the probe for 1 hour.
- 4. Microscope Imaging

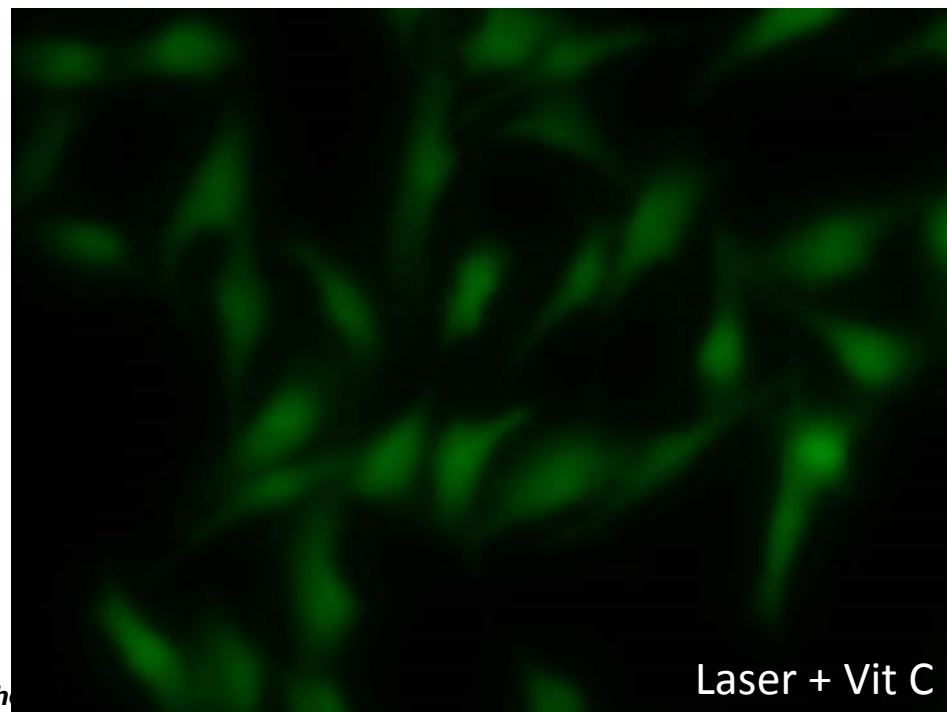
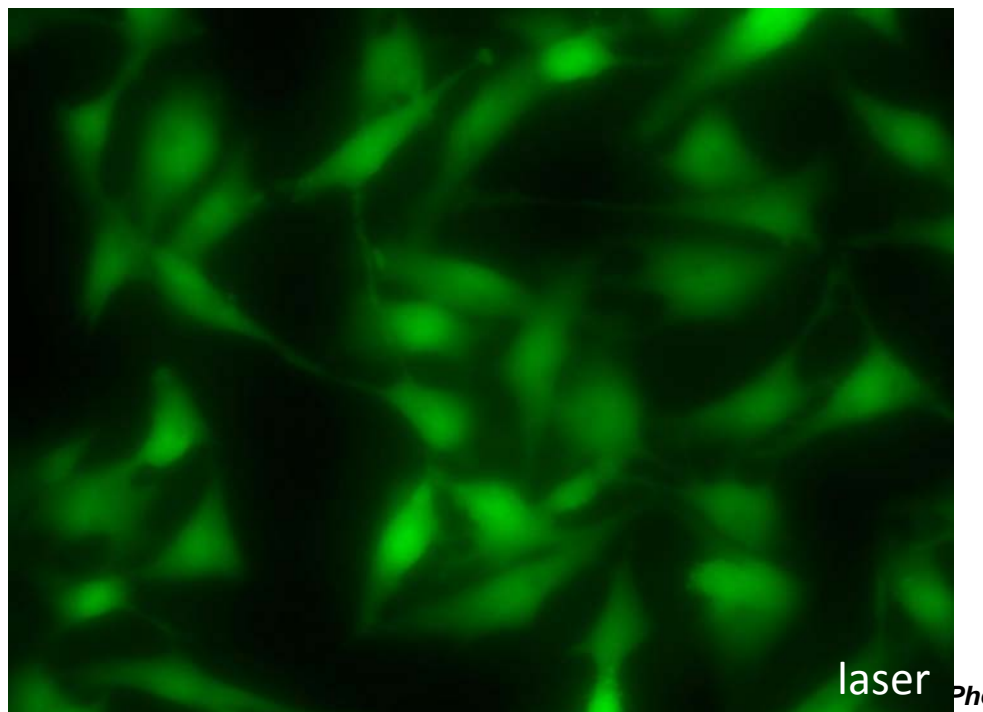
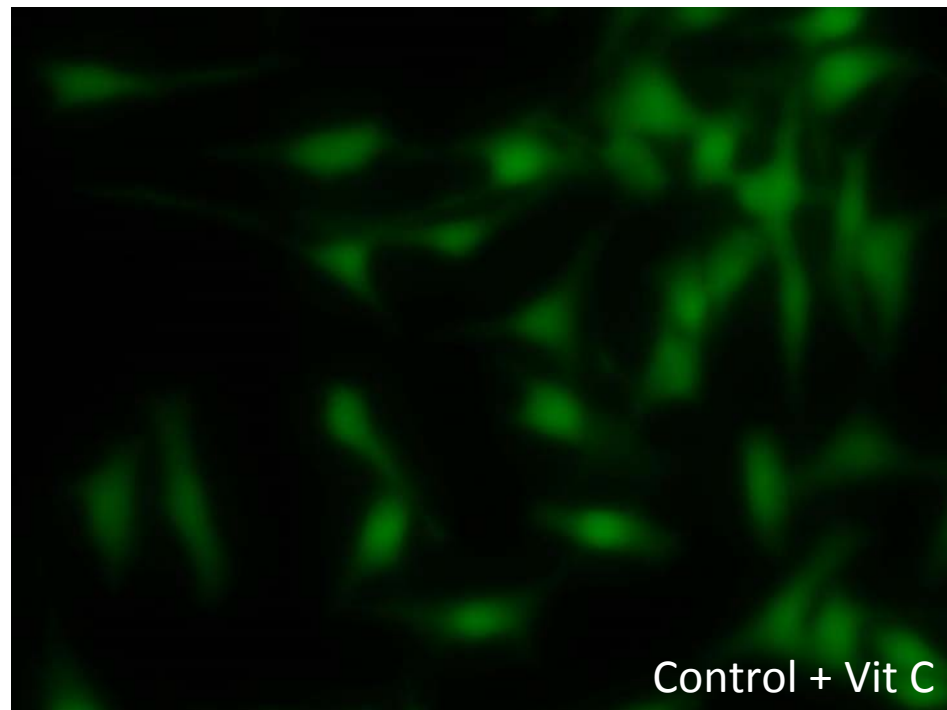
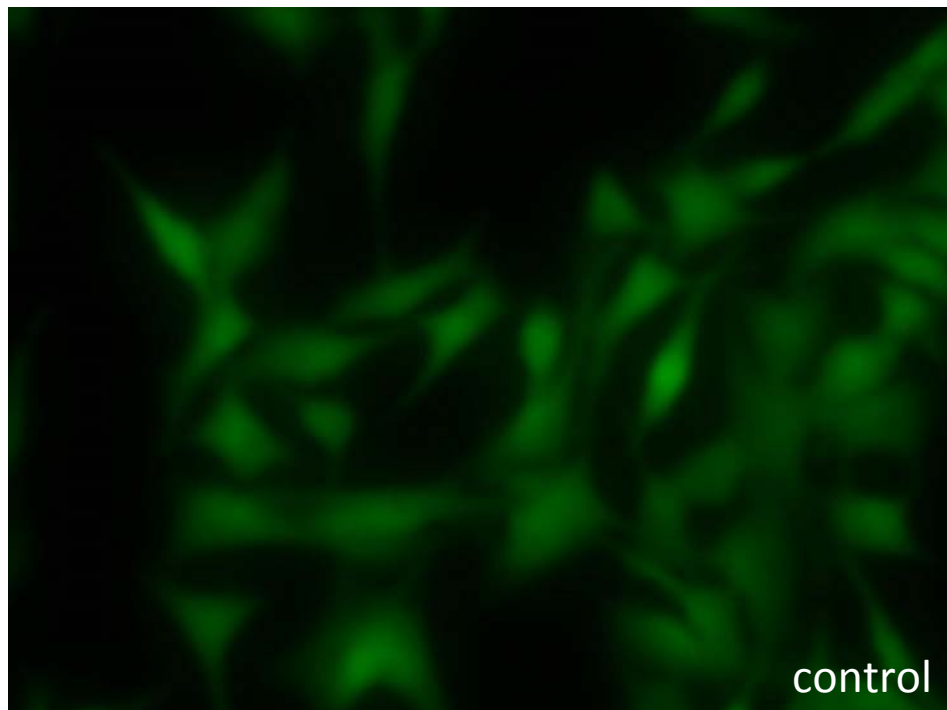


Results

- 1. 810 nm laser promoted ROS production.
- 2. Loading the Probe 30 minutes after the irradiation showed the highest fluorescence.

Ascorbic Acid abrogating induced ROS Production?

- 1. Apply 100 μ M Ascorbic Acid into the medium and pre-incubated for 30 minutes.
- 2. Irradiate the cells. Since the time course showed the best fluorescent results when loading the probe 30 min after irradiation, cells were incubated for 30 minutes before loading the probe.
- 3. 2x washing with PBS and incubate cells in the loading buffer for 1 hour.
- 4. microscopy imaging



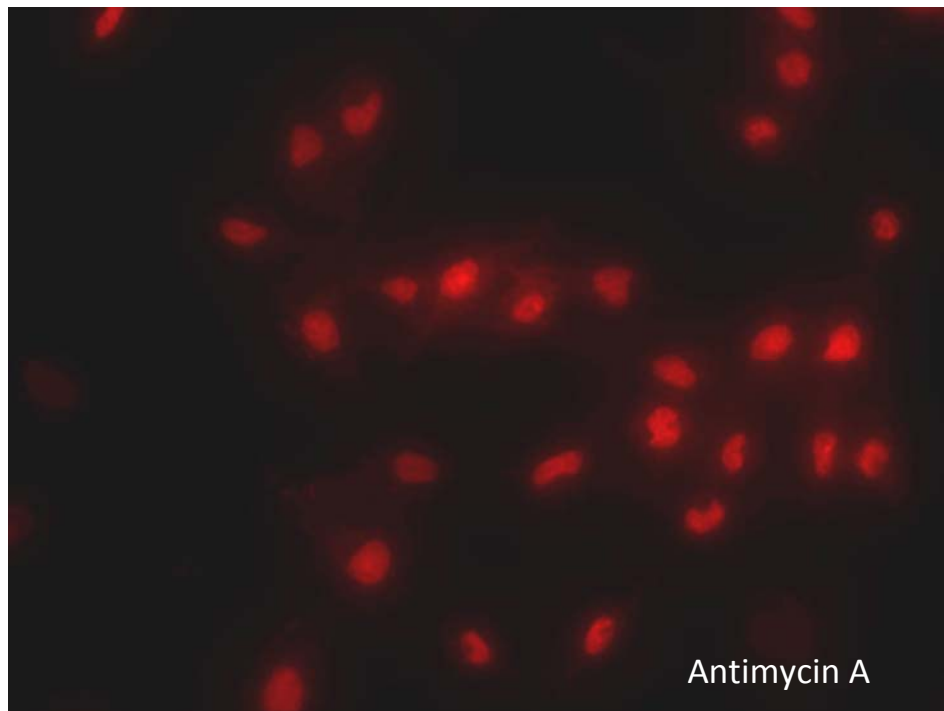
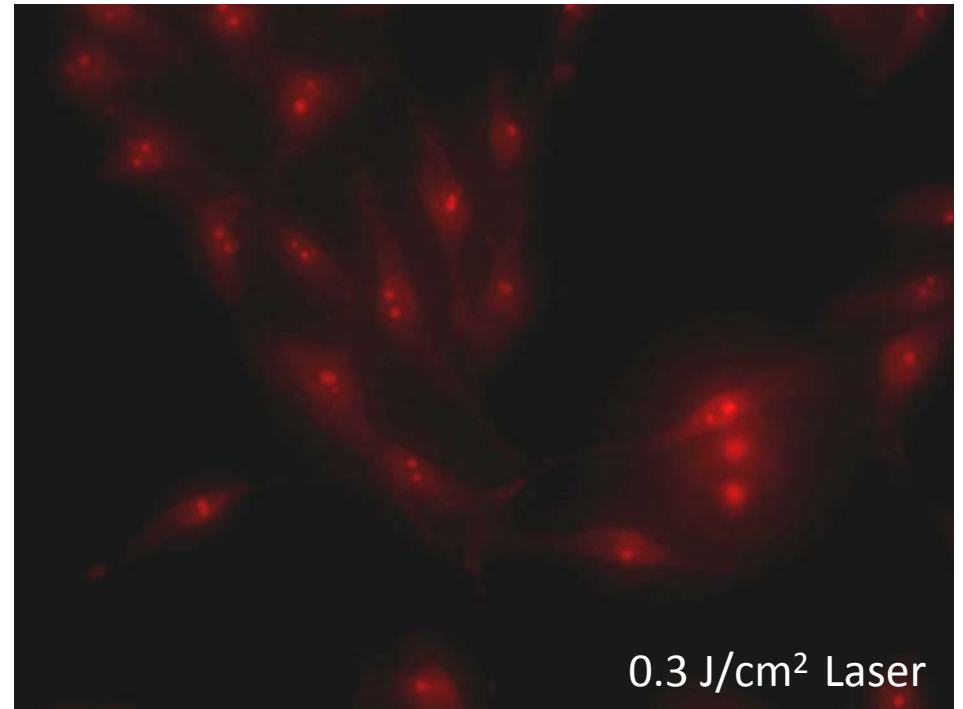
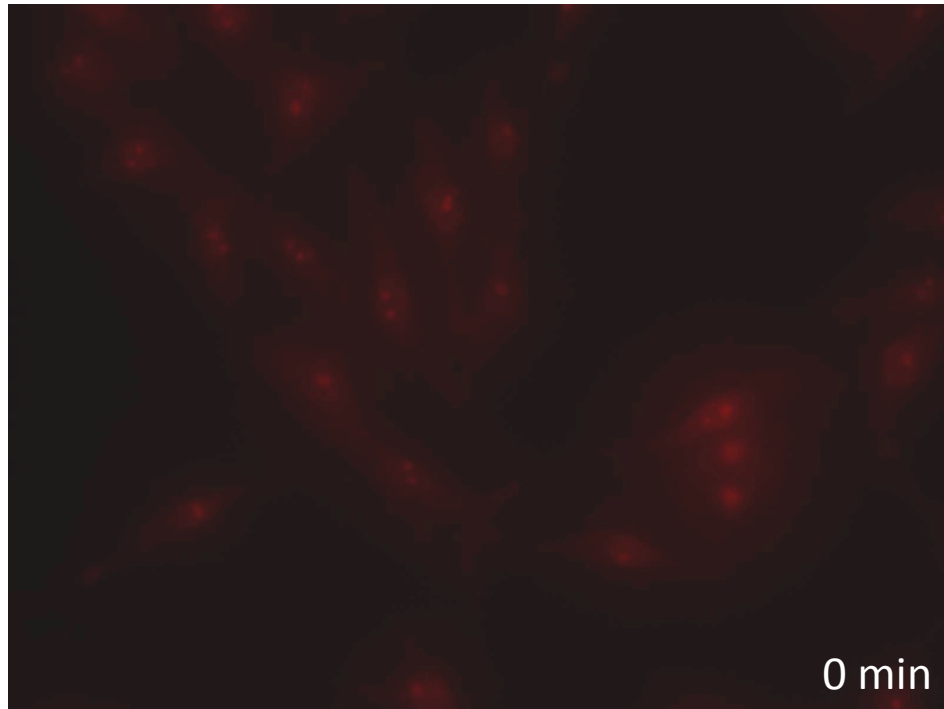
Results

- 1. Ascorbic Acid decreased the DCFDA fluorescence in both control and the laser irradiated cells.

MitoSOX Red Assay

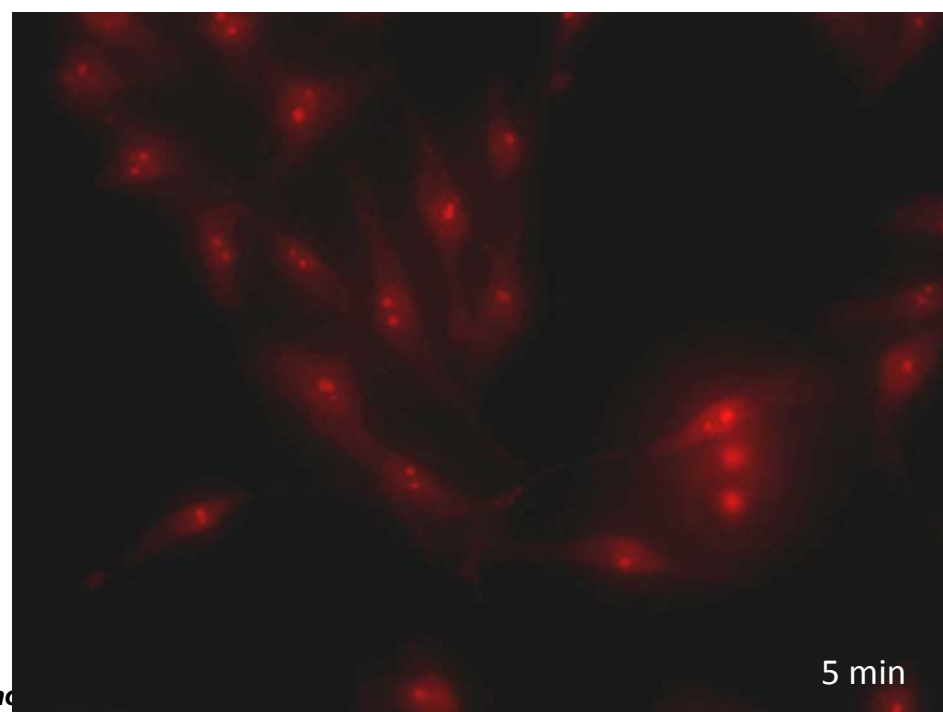
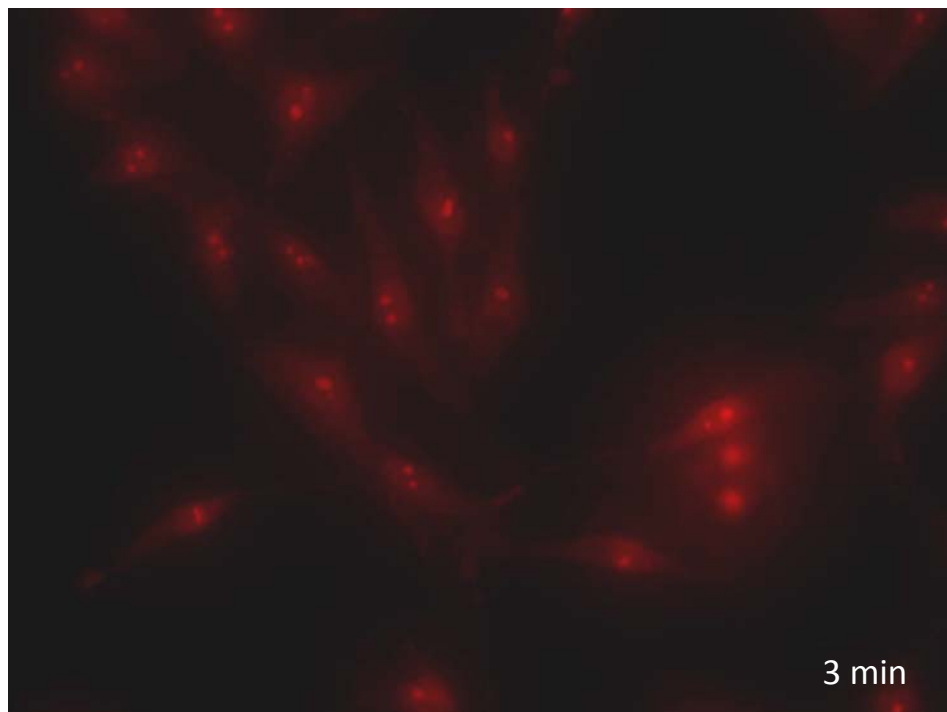
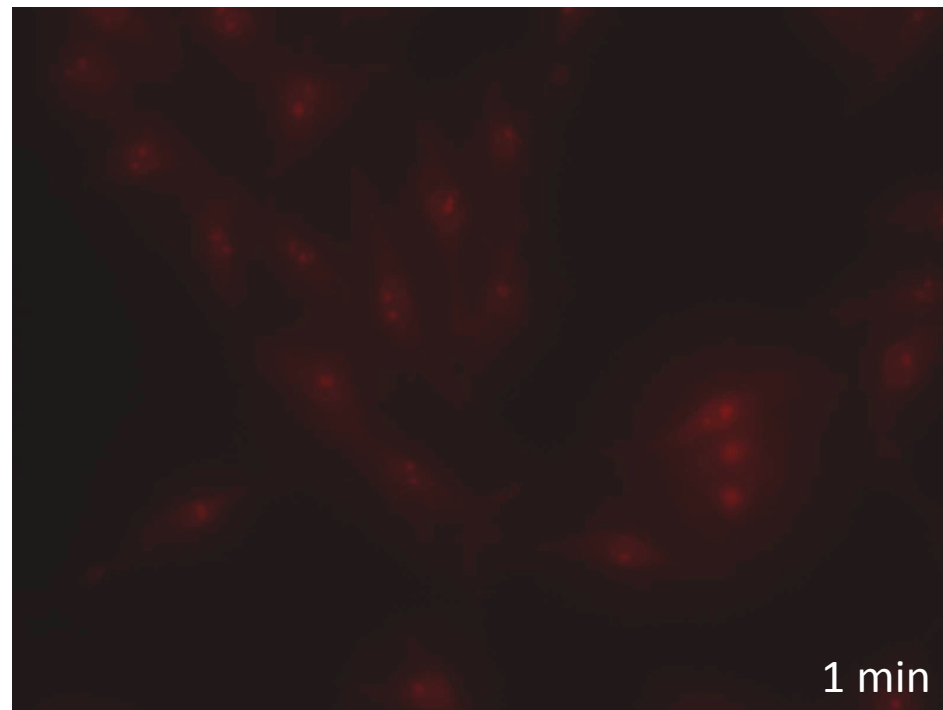
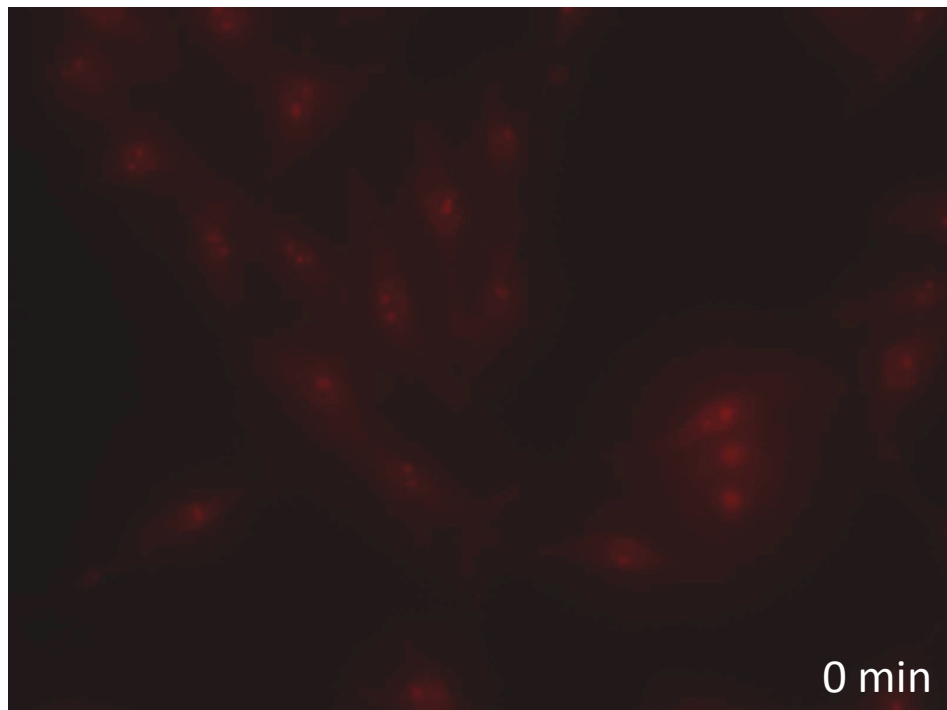
Laser induced superoxide production in the mitochondria.

- 1. Irradiate cells
- 2. 2x washing with PBS and incubated cells in the loading buffer for 1 hour before imaging
- 3. Apply Antimycin A as a positive control



**Mitochondrial Superoxide Production
By
MitoSOX Red**

Real Time Fluorescence of Mitochondrial Superoxide by MitoSOX Red

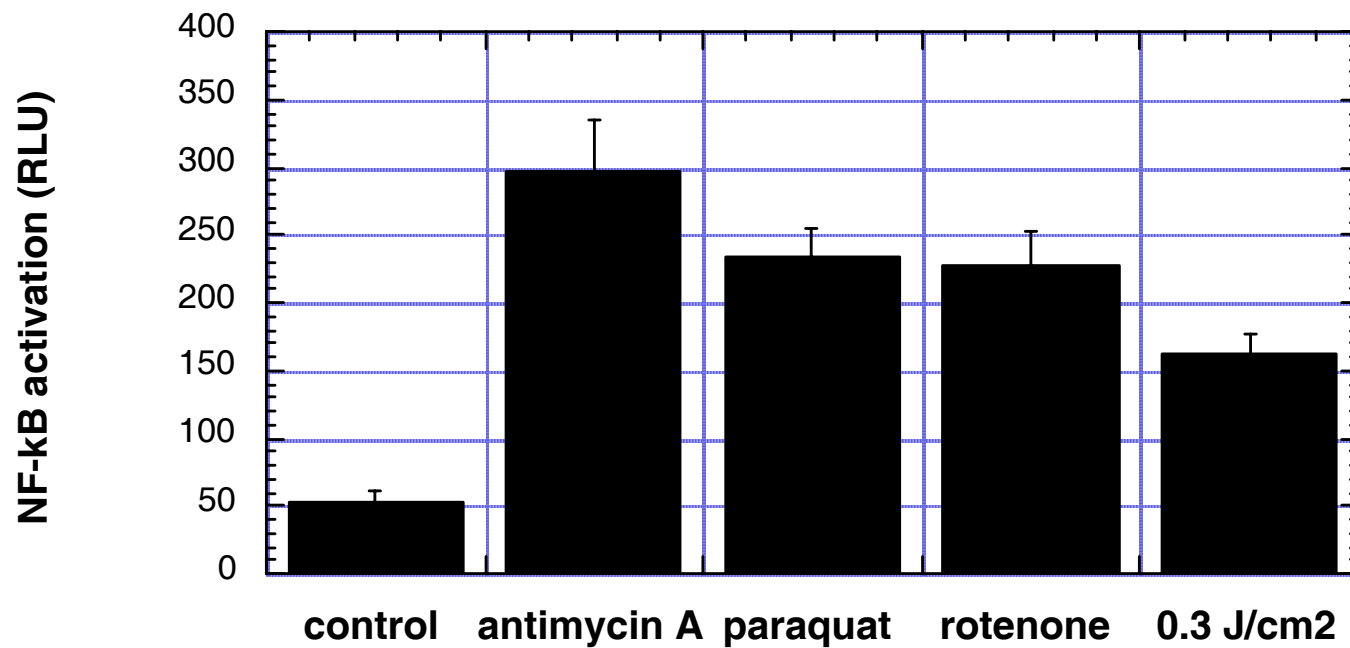


Pho

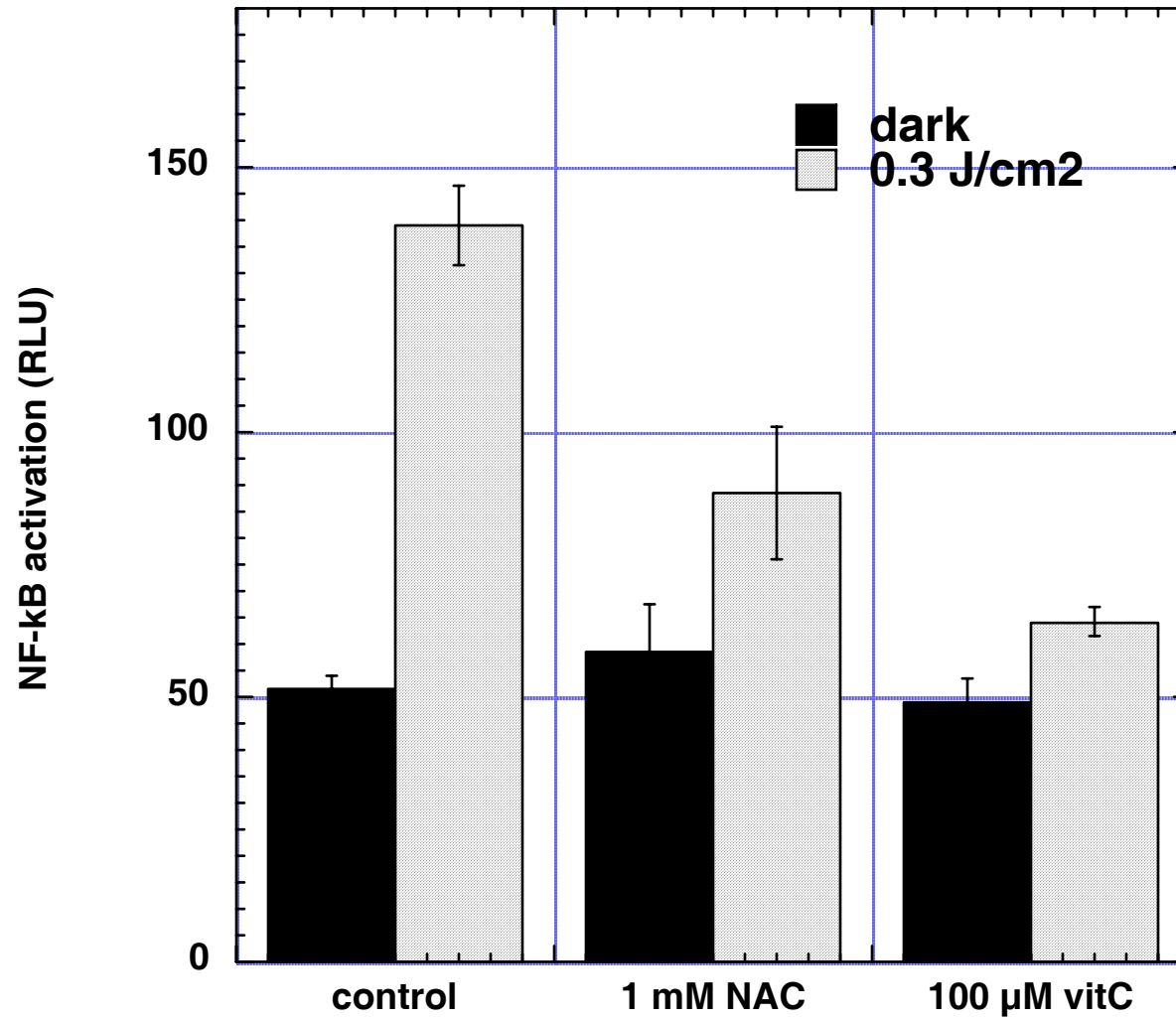
Results

- Fluorescence in the first minute did not show significant difference from the control.
- Fluorescent difference started to appear from the third minutes
- This test serves an additional evidence supporting mitochondrial superoxide was actually produced by 0.3 J/cm^2 laser during the irradiation.

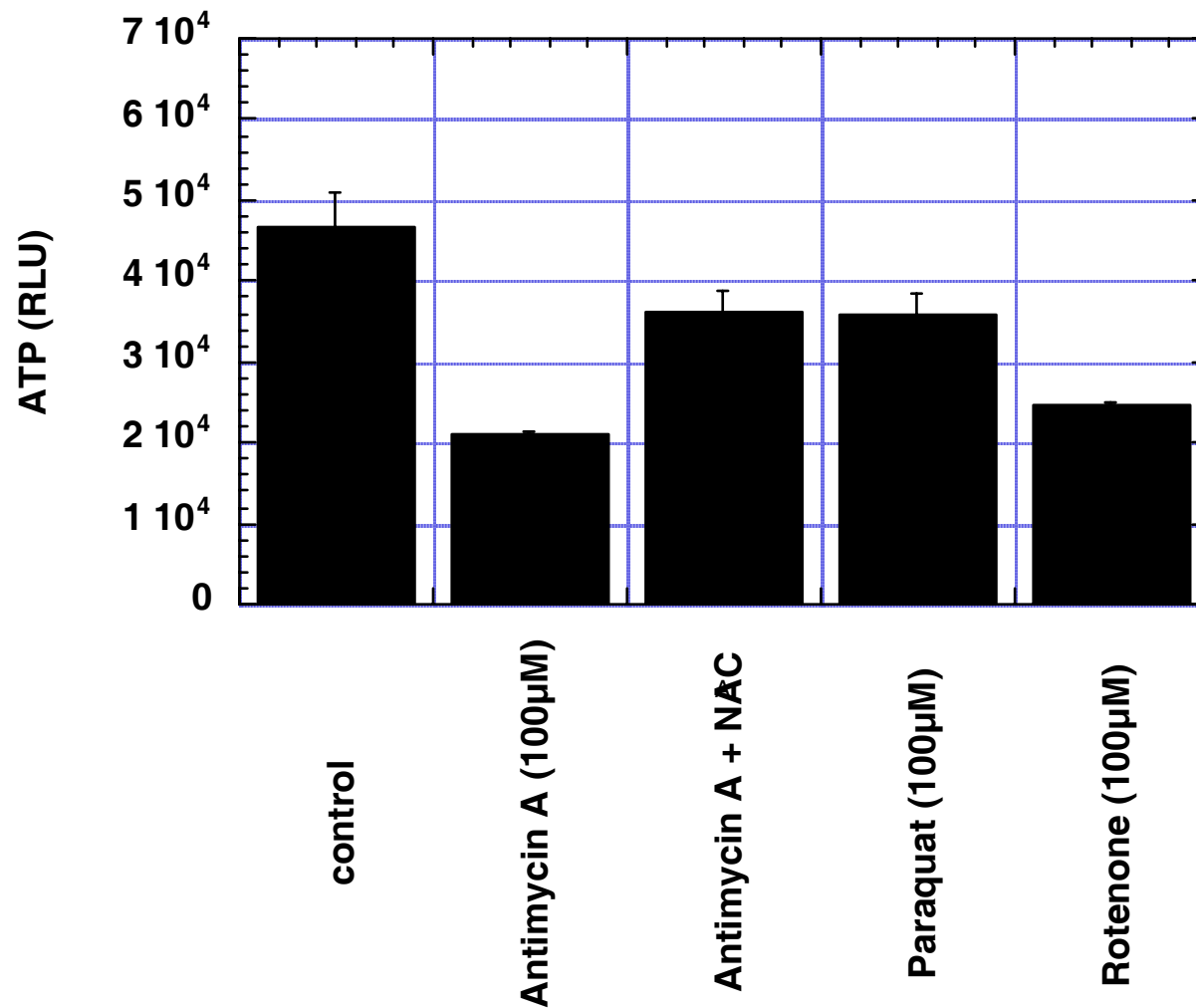
Do mitochondrial inhibitors increase NF-kB activation?



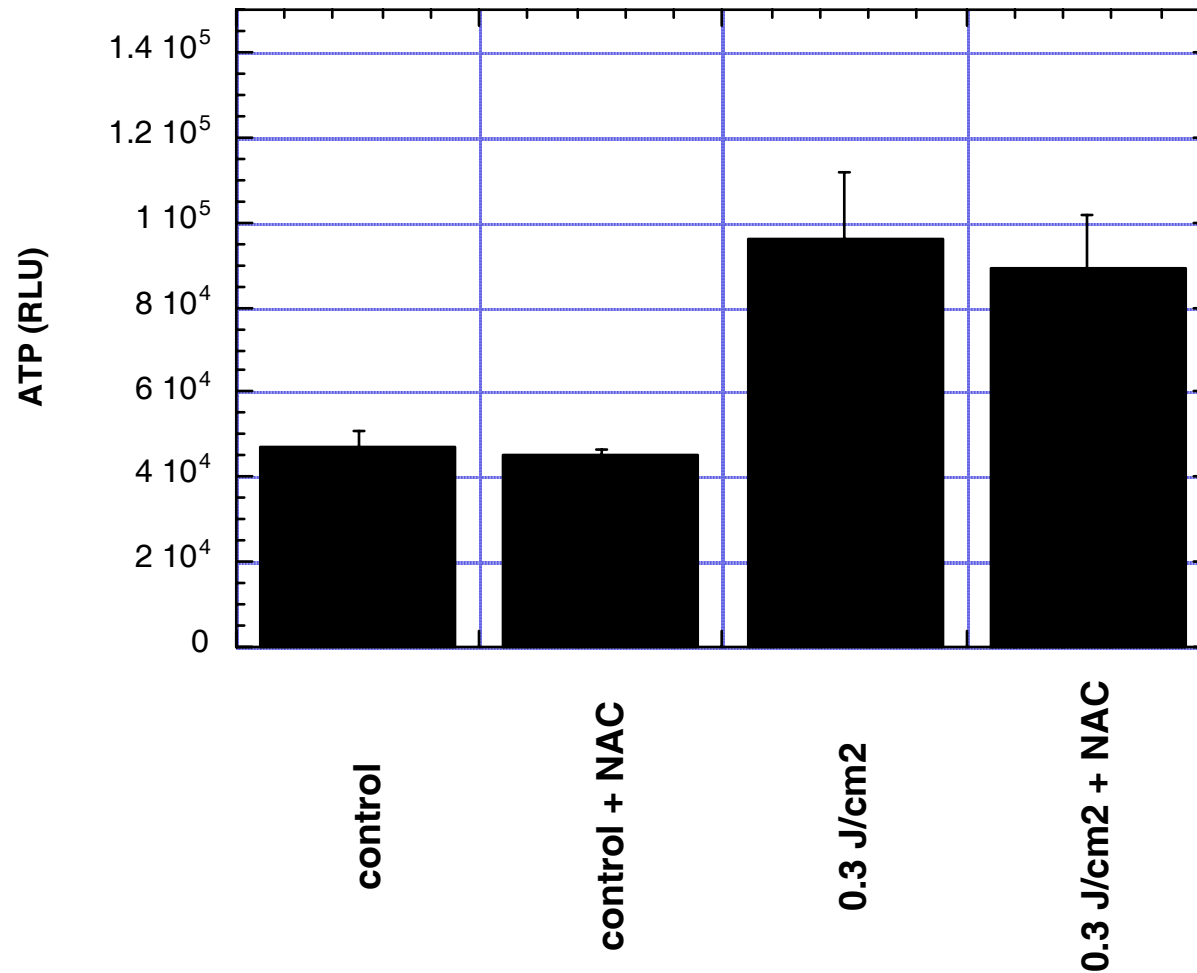
Do antioxidants prevent NF-kB activation?



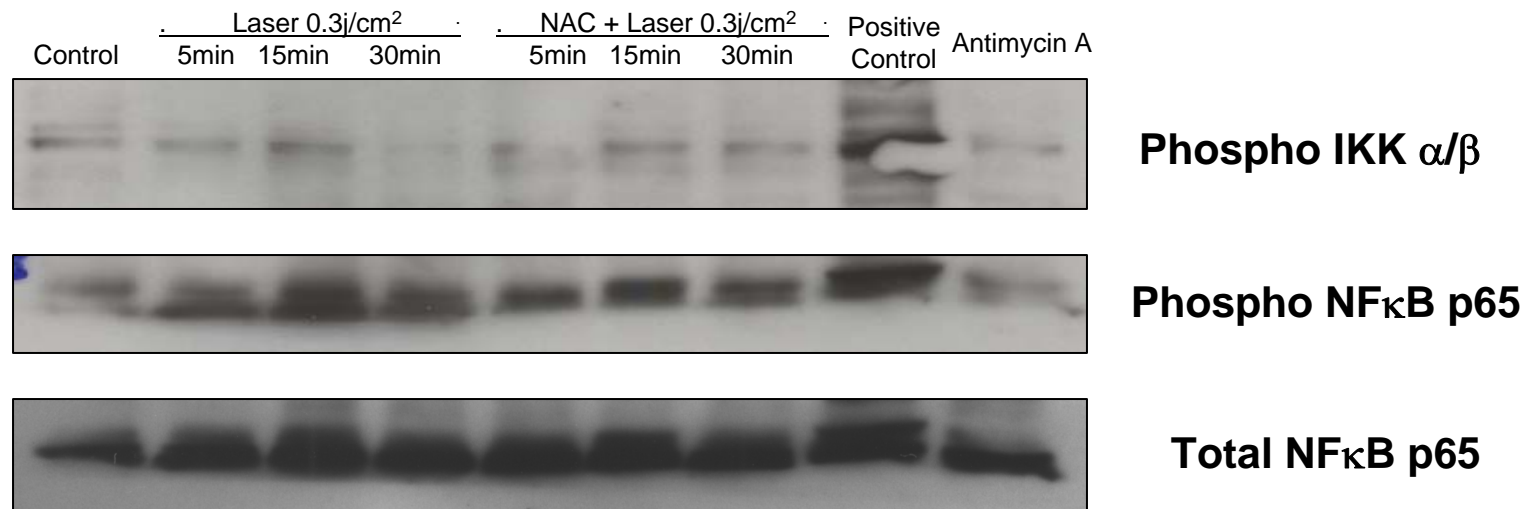
What is effect of mitochondrial inhibitors and antioxidants on ATP?



Do antioxidants abrogate laser induced ATP ?



NF κ B activation by Lasers



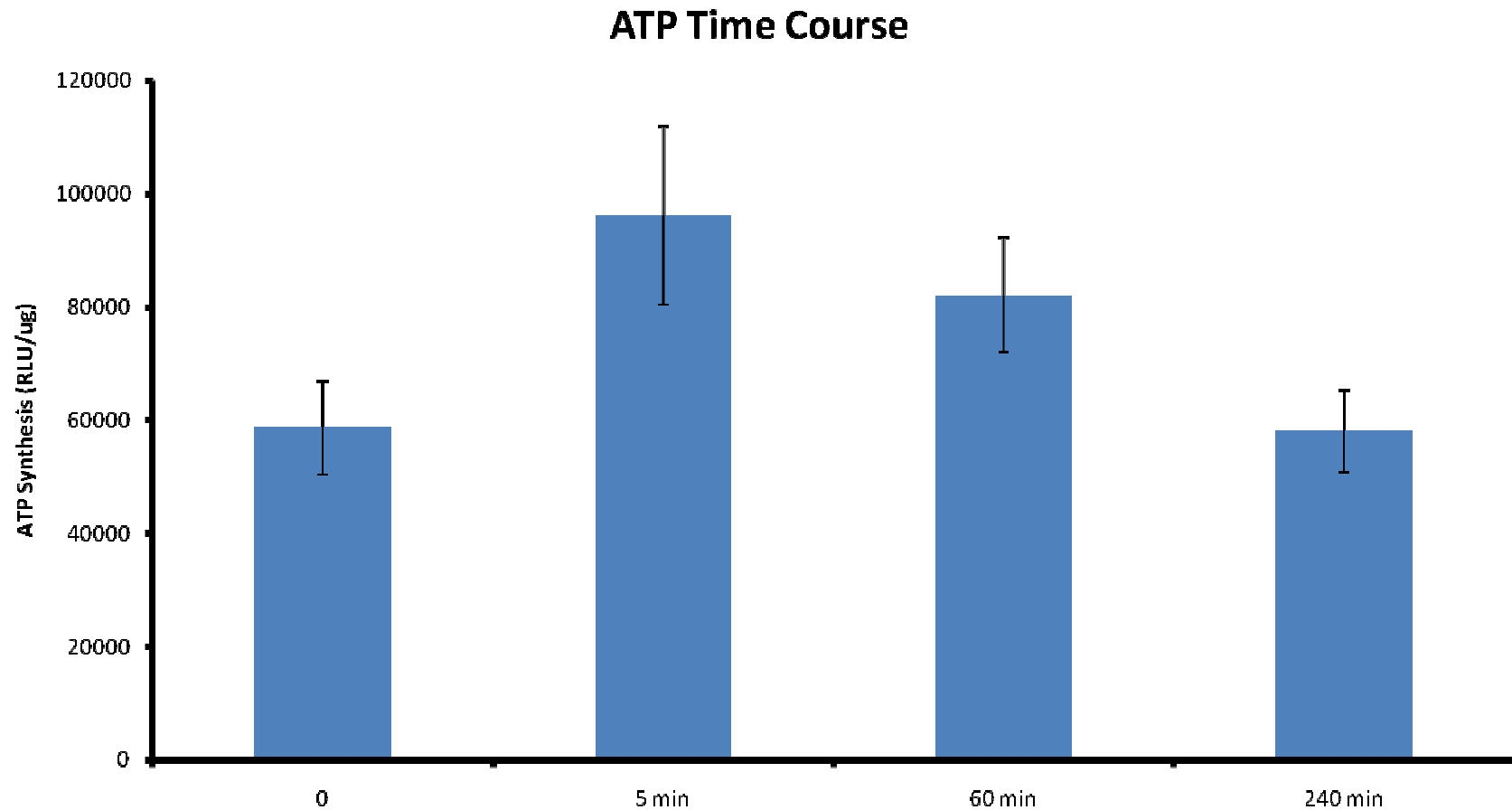
Positive Control: Cells (Primary MEFs) treated with TNF α for 15min

Conclusions: Laser (810nm 0.3j/cm²) can activate NF κ B.

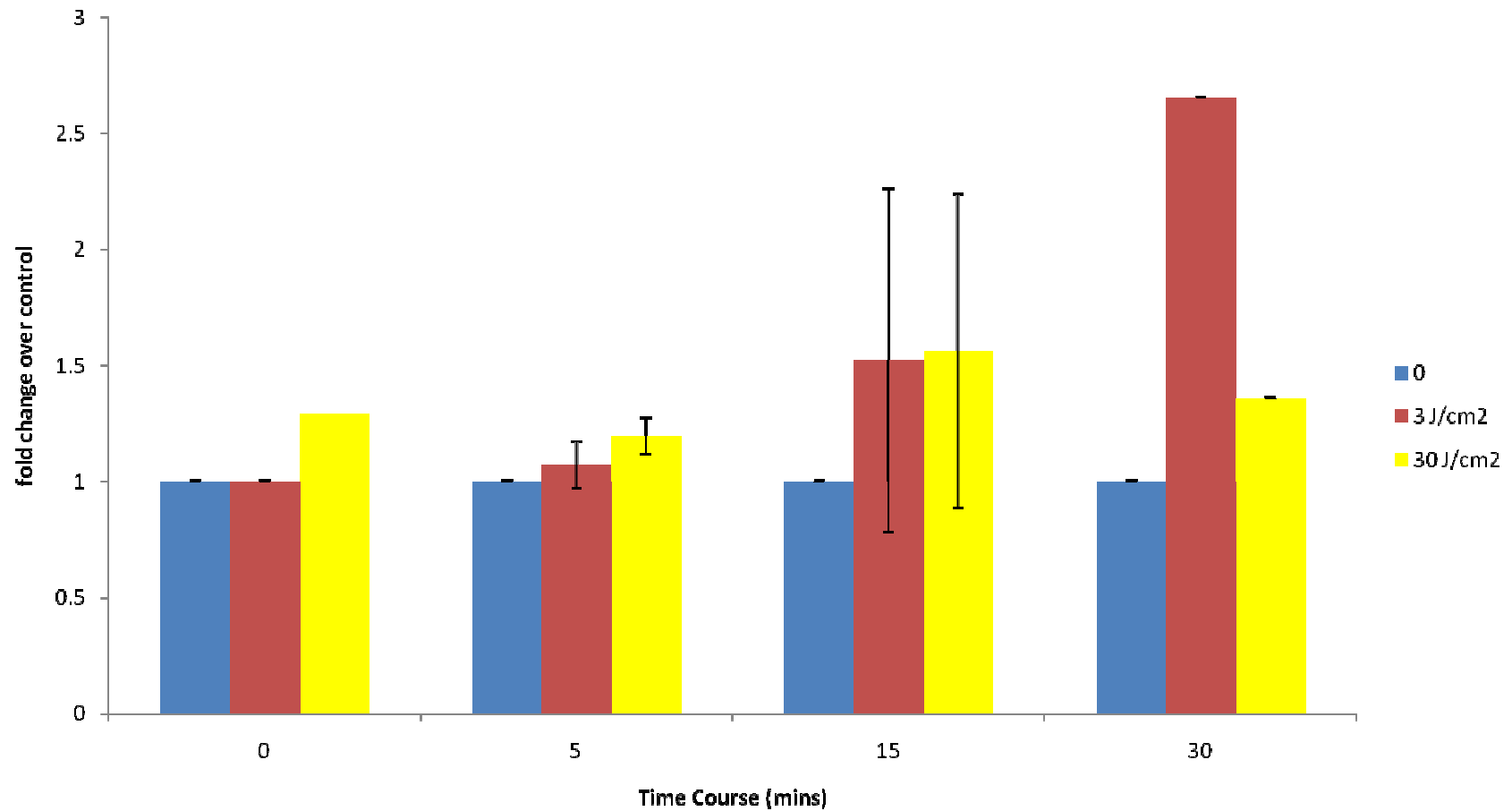
Note: NAC did not neutralize this expression completely suggesting either;

1. incomplete inhibition of ROS
2. alternative (GF) mediated activation of NF κ B.

Human Fibroblasts



810 LED induced Nitrate Production in HeLa

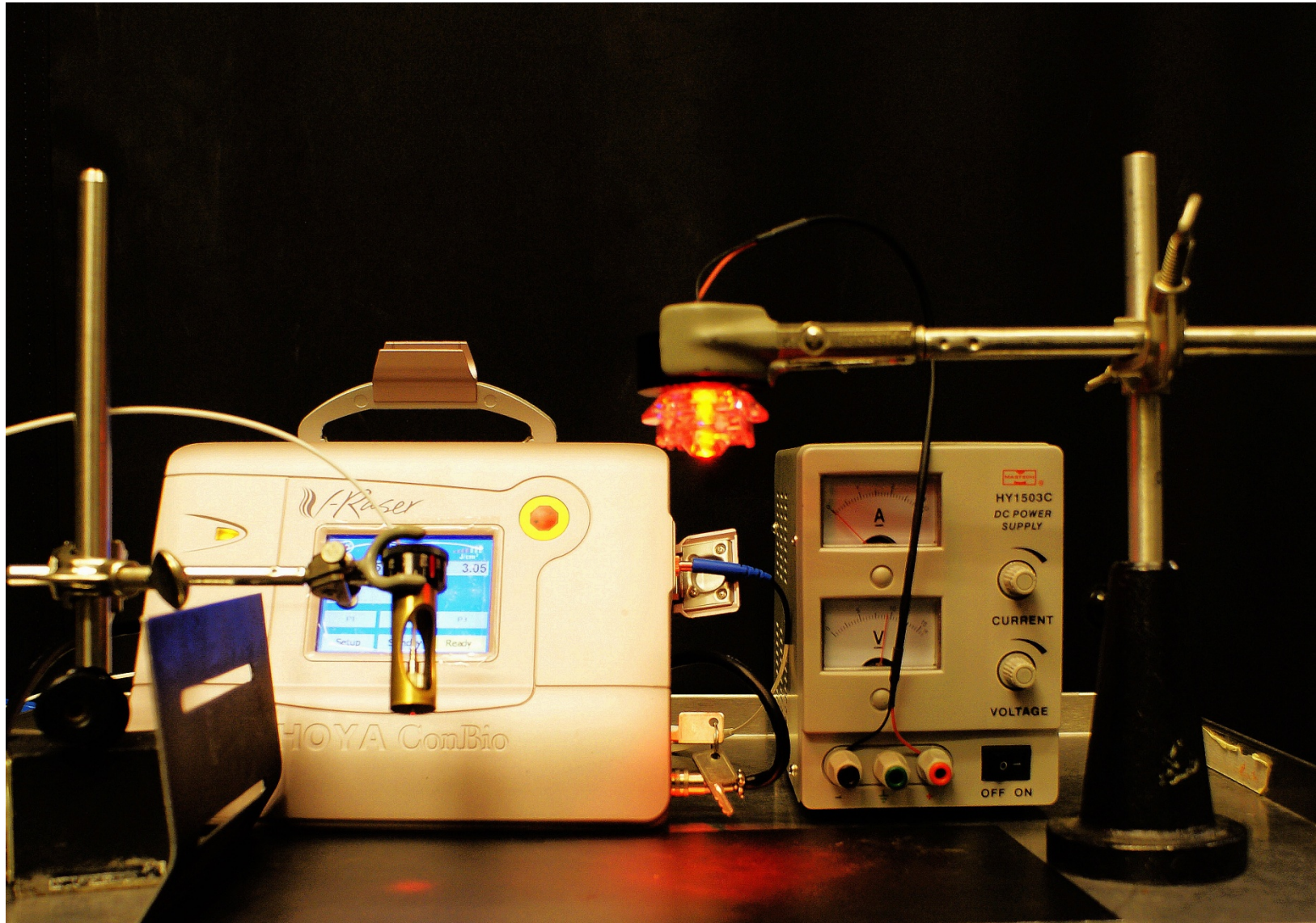


Experiments with HEK293 cells

- Why HEK293?
 - first described in 1977 ([Graham et al., J. Gen Virol 1977 Jul;36\(1\):59-74](#)).
 - Lack of any Toll-like receptors by nature
 - Easy to grow and transfect (transiently and stably)
- Stably transfected TLR9 and NF-kB Luciferase reporter genes into HEK293. (by Marc Lamphier at Eisai.)
- Transiently transfected NF-kB Luciferase reporter
- Irradiated by 810nm LED, 10mW/cm² 5 min.

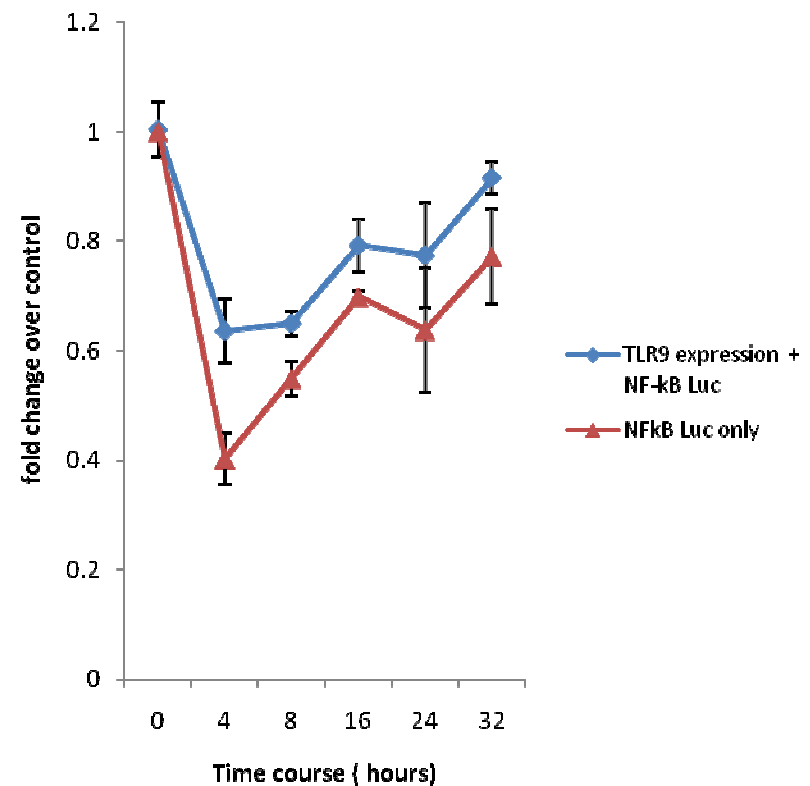
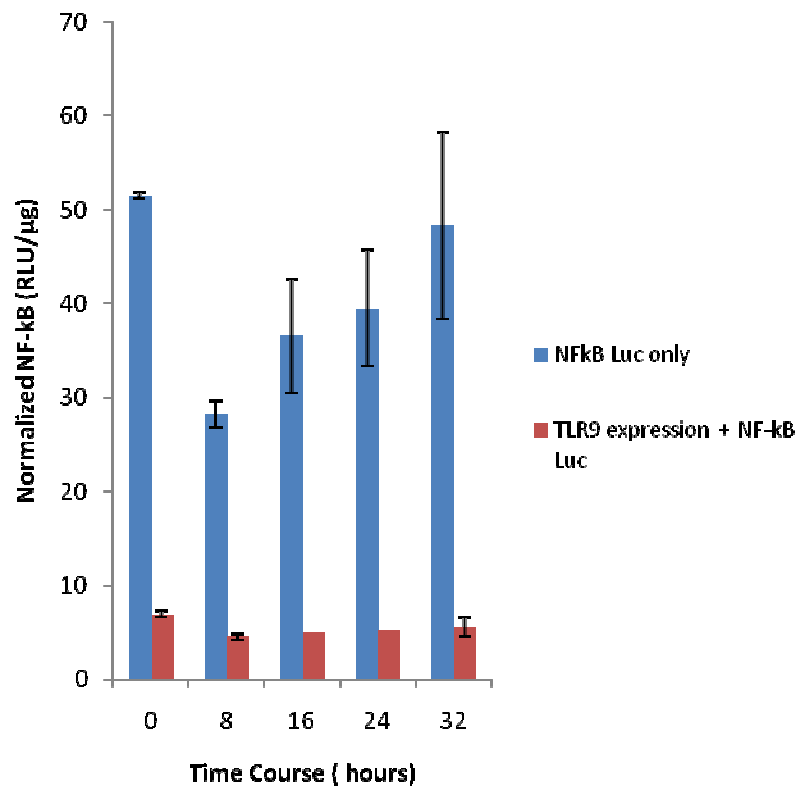
980-nm CW laser

810-nm and 970-nm LED array

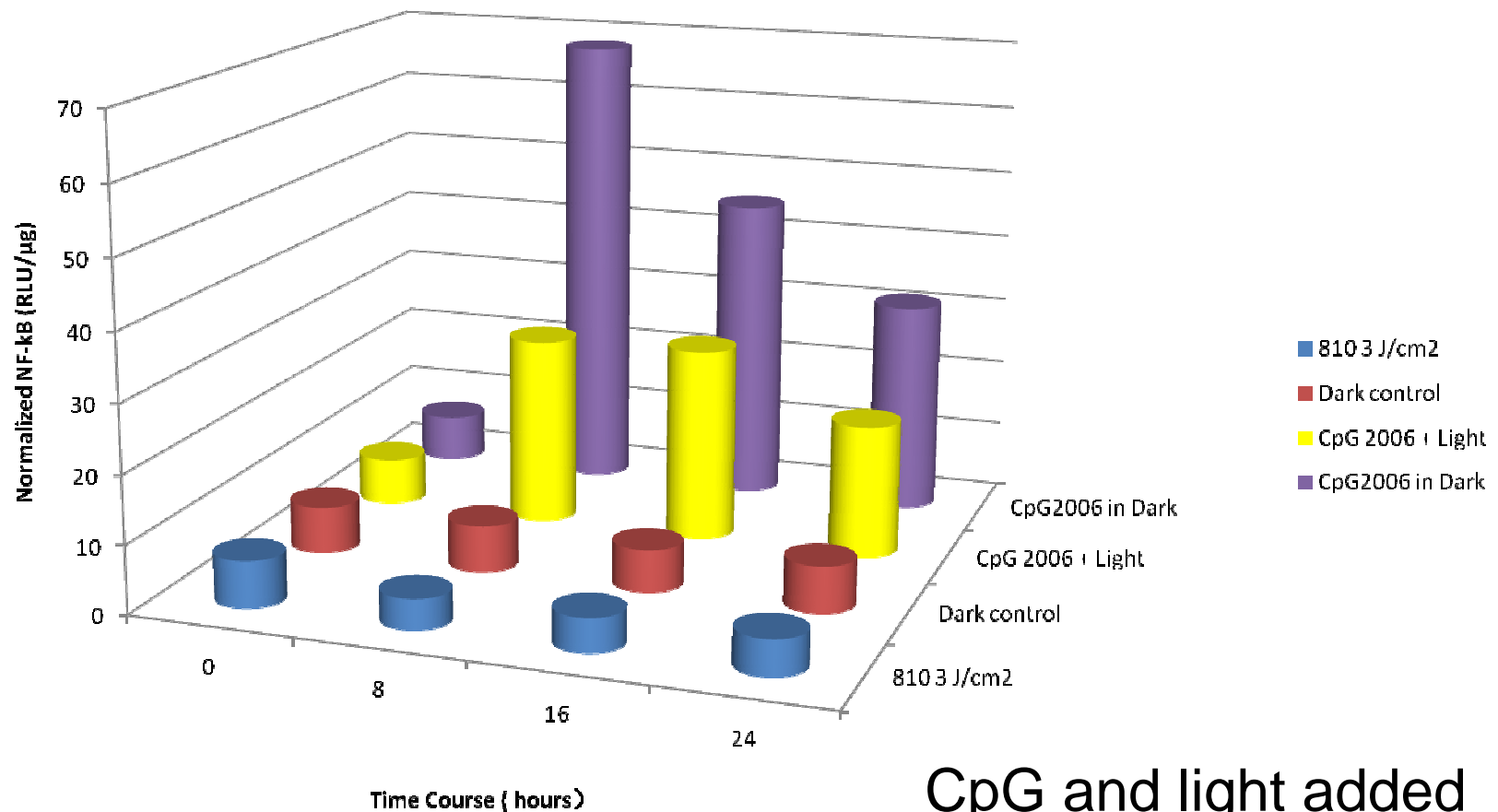


Michael R Hamblin, PhD, Wellman Center for Photomedicine. Dose Response Conference 2009

810 Laser induced NF-kB inhibition in HEK293

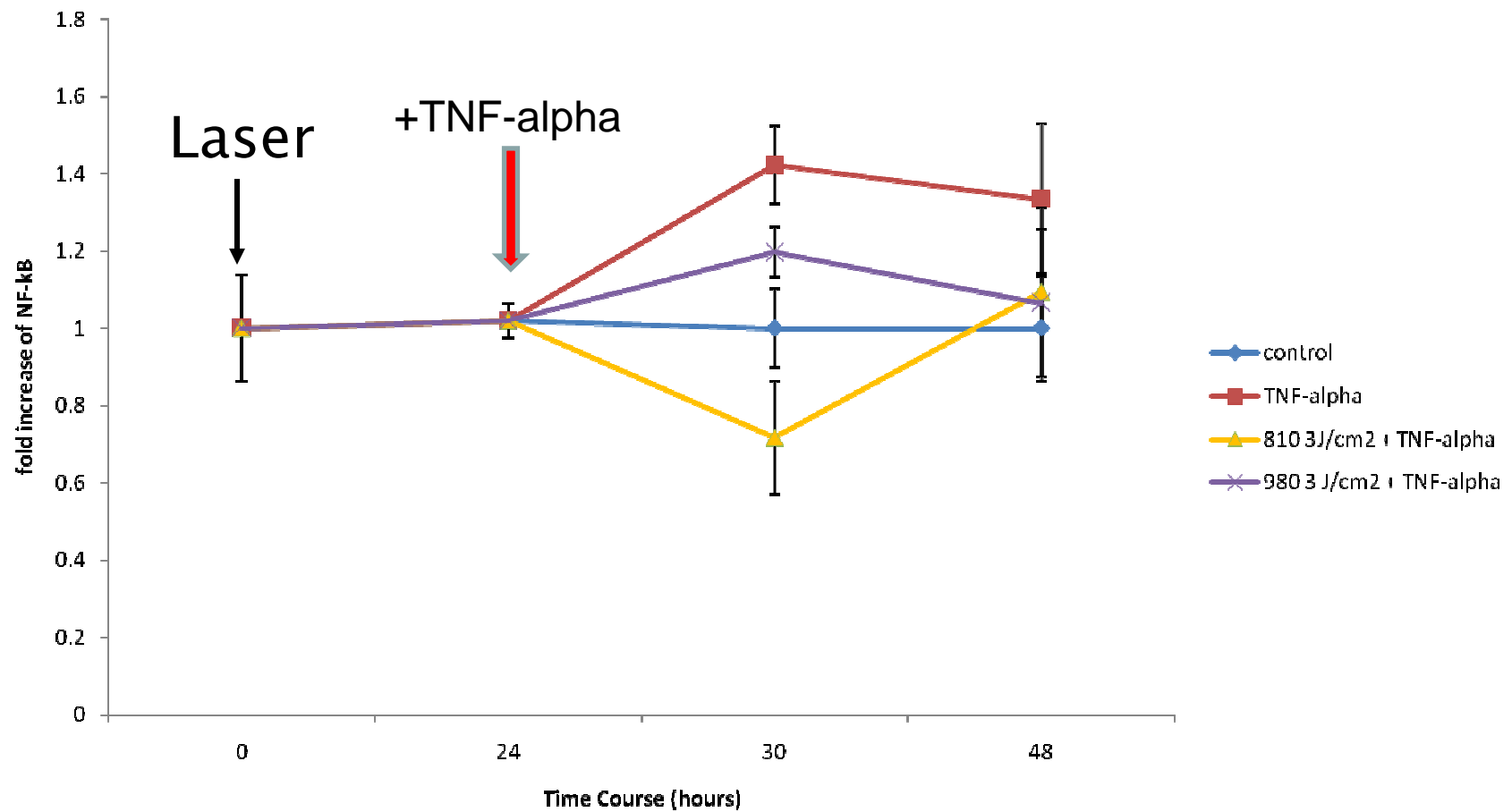


Light effect on TLR9 signaling

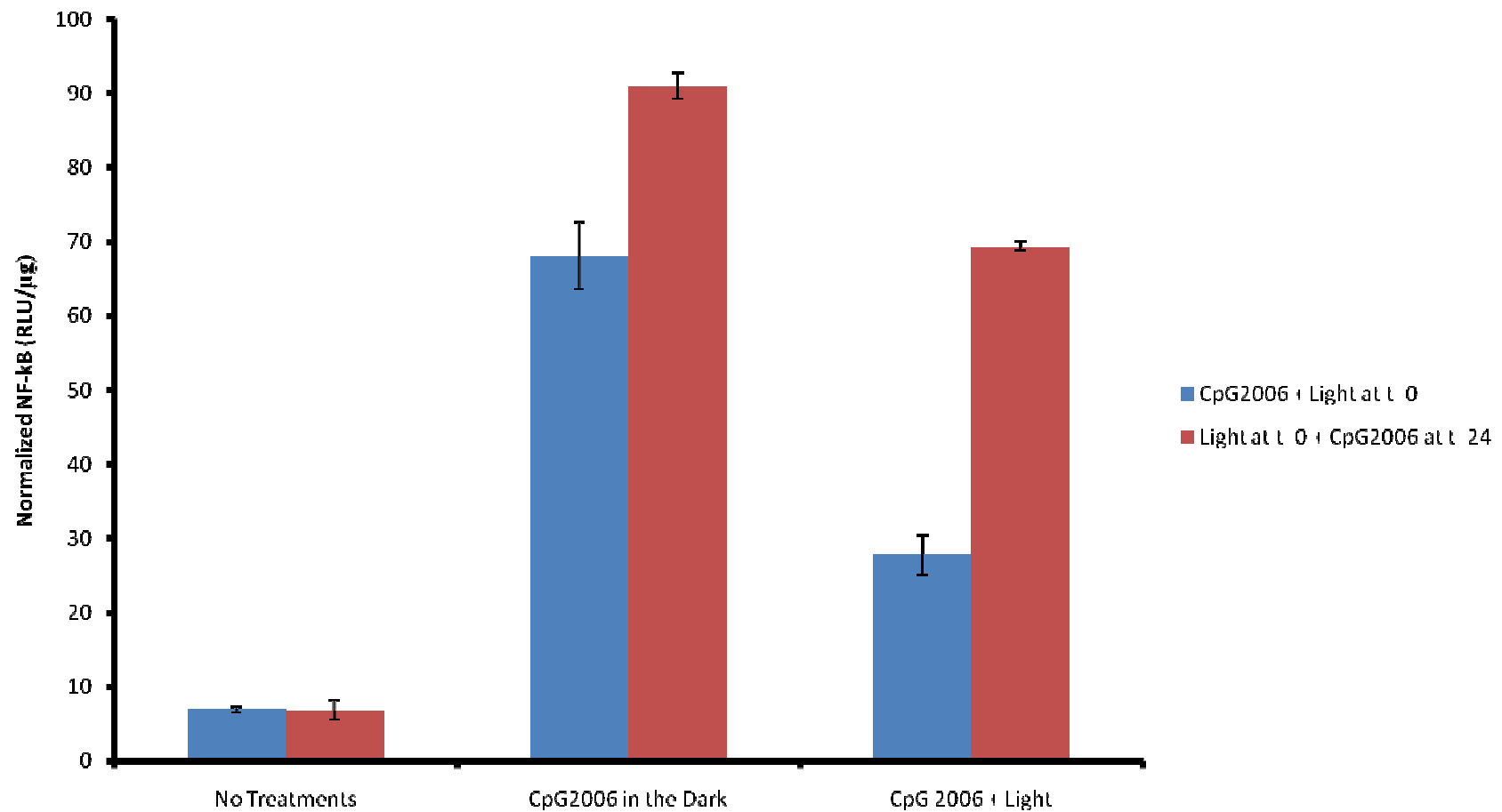


CpG and light added
at same time

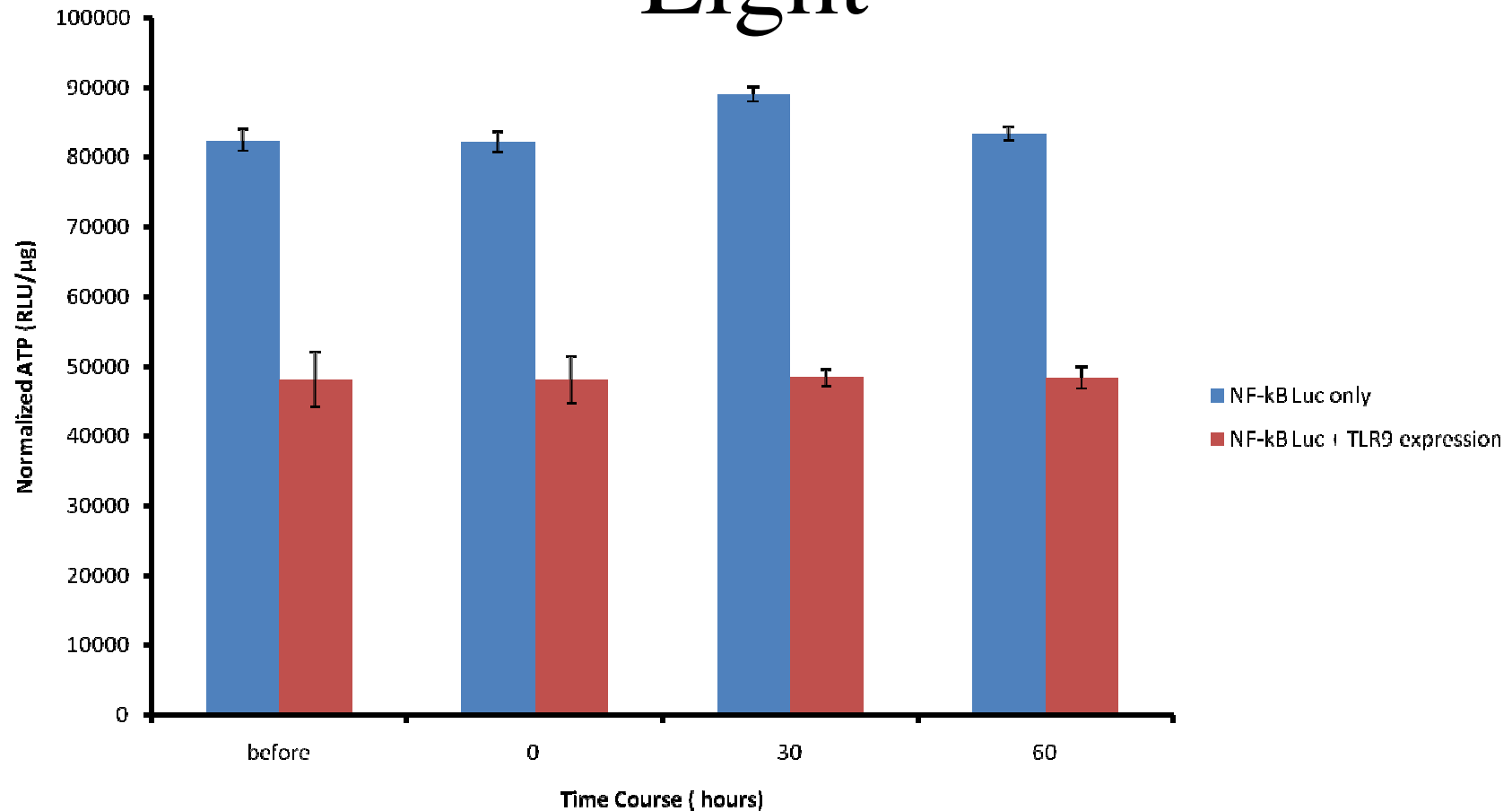
Laser pre-illumination abrogates effects of TNF-alpha 24 h later



Light effect on TLR9 Signaling

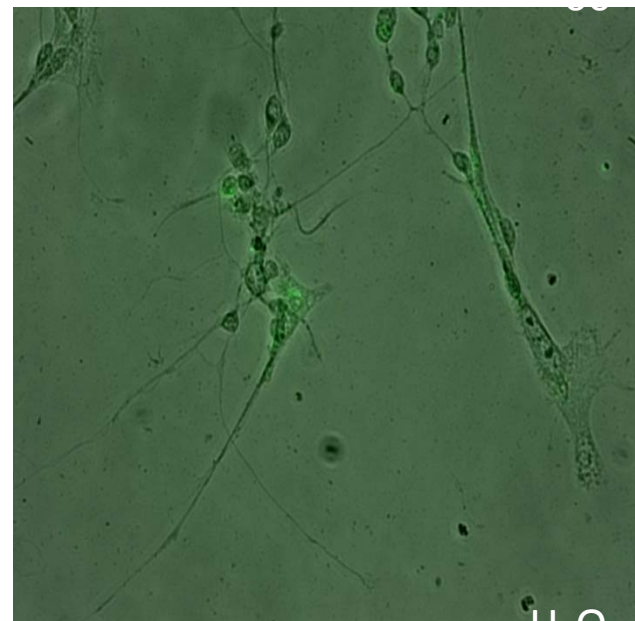
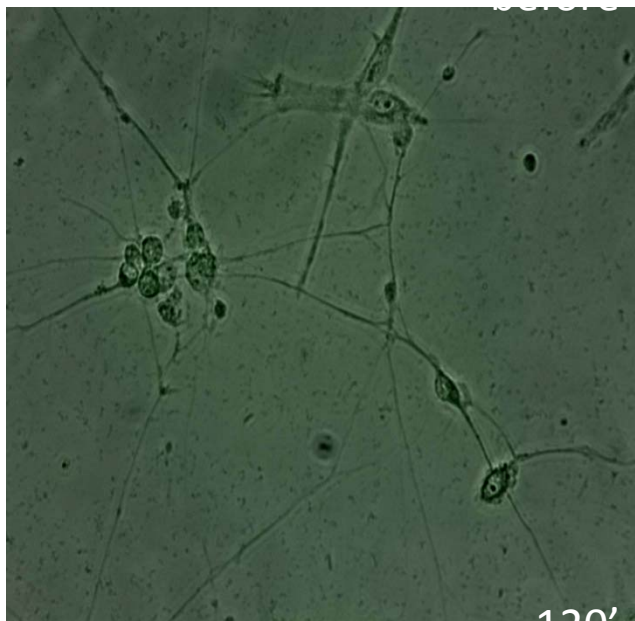
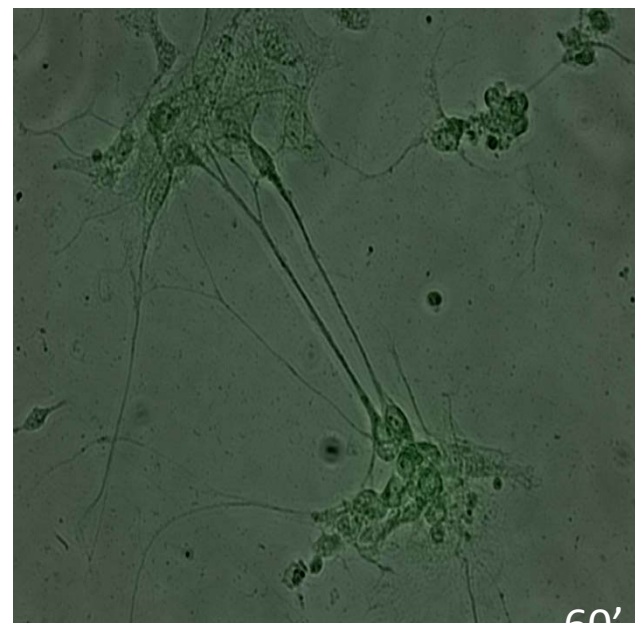
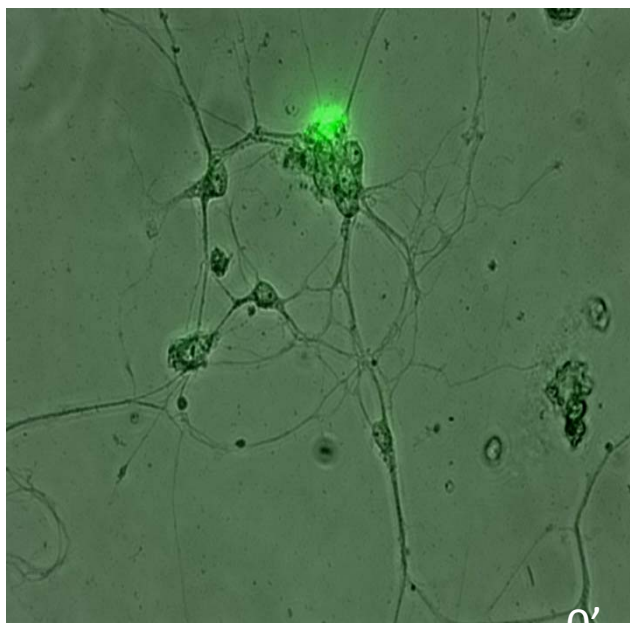
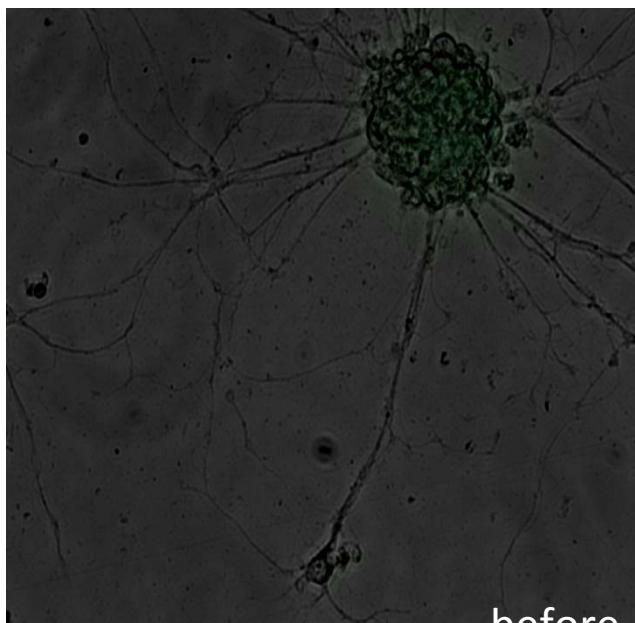


ATP change by 810nm 3J/cm² Light

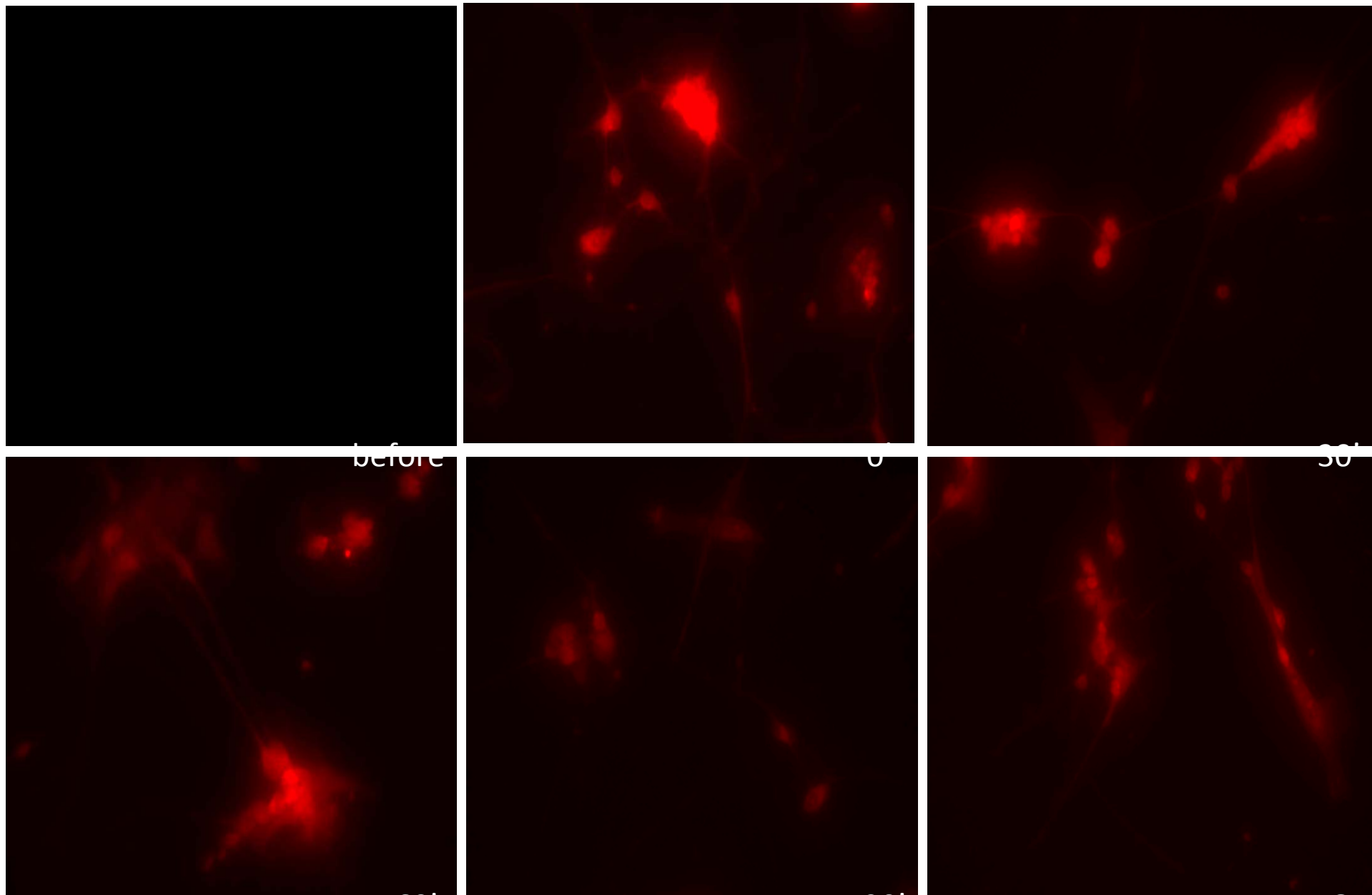


ROS in Murine Embryonic Neurons

DCDHF-DA



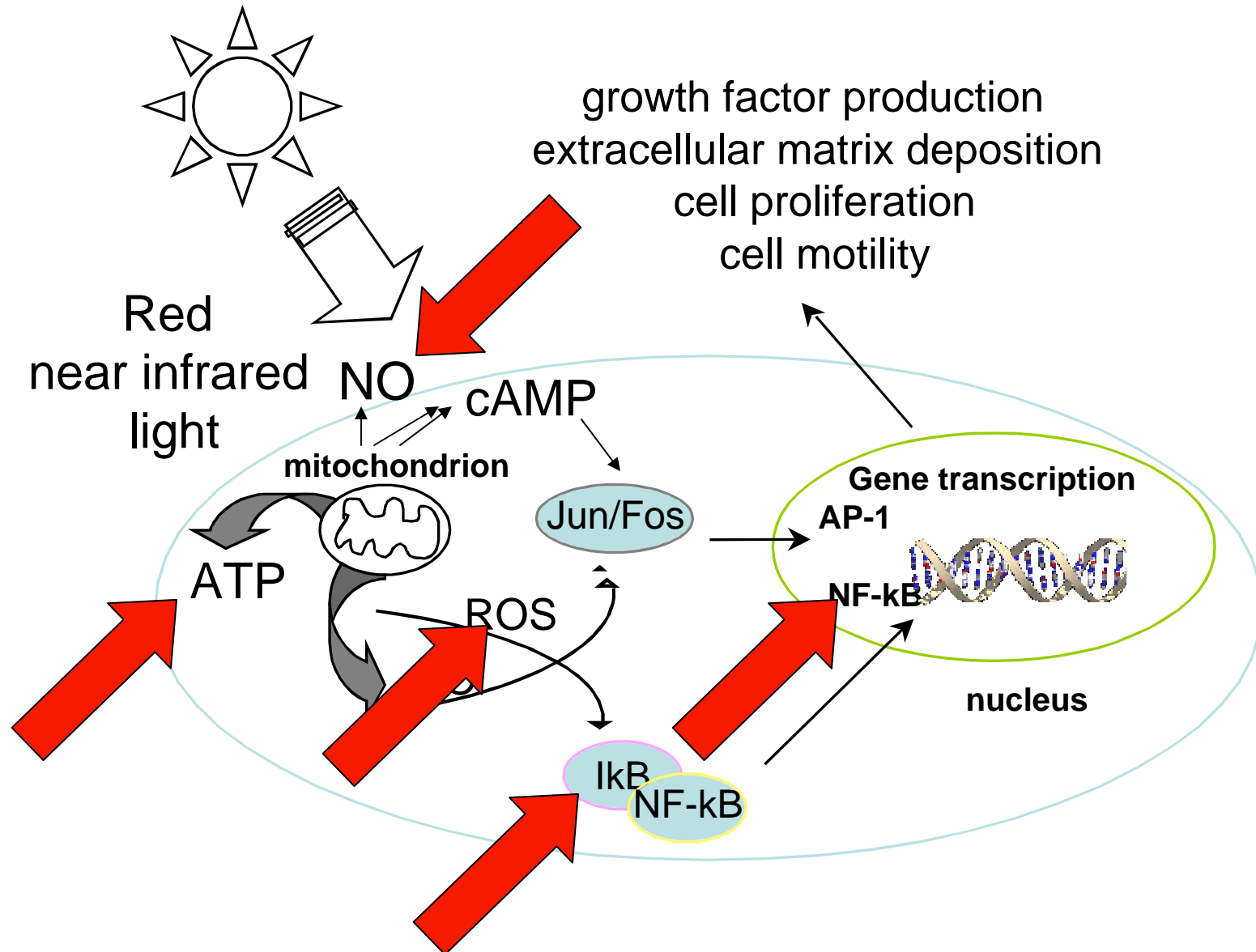
MitoSox Red



Conclusions I

- 1 810-nm laser increases cellular ATP
- 2 810-nm laser activates NF- κ B
- 3 810-nm produced mitochondrial ROS
- 4 Mitochondrial inhibitors and (H_2O_2) activate NF- κ B without increasing ATP
- 5 Mitochondrial inhibitors produce ROS
- 6 Antioxidants abrogate laser and mitochondrial inhibitor-induced NF- κ B activation but have no effect on ATP
- 7 Preliminary evidence of NO production 15 and 30 min post-light
- 8 Different cell types (HEK293 cells and neurons) may behave differently

Mechanisms of LLLT



What is the mechanism for biphasic dose response?

ROS have been shown many times to stimulate at low doses but to be harmful at high doses

NO (and peroxynitrite) may also have biphasic response: stimulate at low dose and inhibit at high dose

Protective transcription factors may be induced at low dose (NF- κ B) and additional different harmful pathways activated at high dose

Future work

Test other cell types: neurons, leukocytes, epithelial cells

Test other wavelengths and non-coherent light sources

Investigate other redox sensitive transcription factors (AP1)

Repeat experiments (bioluminescence imaging, effect of antioxidants) in vivo

Study mouse model of traumatic brain injury

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